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3 Invited Review

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5 **The flagellum of *Trypanosoma brucei*: new tricks from an old dog**

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9 Katherine S. Ralston<sup>a</sup>, Kent L. Hill<sup>a,b,\*</sup>

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17 <sup>a</sup>*Department of Microbiology, Immunology, and Molecular Genetics, University of California, Los*  
18 *Angeles, CA 90095 USA*

19 <sup>b</sup>*Molecular Biology Institute, University of California, Los Angeles, CA 90095 USA*  
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29 \*Corresponding author.

30 Kent L. Hill, Ph.D., Department of Microbiology, Immunology and Molecular Genetics, University of  
31 California, Los Angeles , 609 Charles E. Young Drive,

32 Los Angeles, CA 90095 USA.

33 Tel.: +1-310-267-0546; Fax: +1-310-206-5231.

34 *E-mail address:* kenthill@mednet.ucla.edu  
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## 1   **Abstract**

2           African trypanosomes, i.e. *Trypanosoma brucei* and related sub-species, are devastating human  
3   and animal pathogens that cause significant human mortality and limit sustained economic  
4   development in sub-Saharan Africa. *Trypanosoma brucei* is a highly motile protozoan parasite and  
5   coordinated motility is central to both disease pathogenesis in the mammalian host and parasite  
6   development in the tsetse fly vector. Since motility is critical for parasite development and  
7   pathogenesis, understanding unique aspects of the *T. brucei* flagellum may uncover novel targets for  
8   therapeutic intervention in African sleeping sickness. Moreover, studies of conserved features of the *T.*  
9   *brucei* flagellum are directly relevant to understanding fundamental aspects of flagellum and cilium  
10   function in other eukaryotes, making *T. brucei* an important model system. The *T. brucei* flagellum  
11   contains a canonical 9 + 2 axoneme, together with additional features that are unique to kinetoplastids  
12   and a few closely-related organisms. Until recently, much of our knowledge of the structure and  
13   function of the trypanosome flagellum was based on analogy and inference from other organisms.  
14   There has been an explosion in functional studies in *T. brucei* in recent years, revealing conserved as  
15   well as novel and unexpected structural and functional features of the flagellum. Most notably, the  
16   flagellum has been found to be an essential organelle, with critical roles in parasite motility,  
17   morphogenesis, cell division and immune evasion. This review highlights recent discoveries on the *T.*  
18   *brucei* flagellum.

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20   Keywords: Flagellum; Cilium; Motility; Axoneme; Trypanosome; Dynein; Cytokinesis

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## 1    **1. Introduction**

2            The eukaryotic flagellum is an evolutionarily “ancient” organelle. Flagella are represented in  
3    most eukaryotic lineages, from deep branching eukaryotes, e.g. *Giardia*, trichomonads and  
4    kinetoplastids, to crown eukaryotes, e.g. humans and other mammals. In all cases flagella have a  
5    conserved underlying structure and are usually assembled via the conserved process of intraflagellar  
6    transport (Rosenbaum, 2002). Thus, the combined evidence indicates that the flagellum arose very  
7    early in eukaryotic evolution and that organisms lacking flagella arose through loss of the organelle.  
8    The flagellum is also “old” as an object of study and is arguably the first organelle ever described  
9    (Satir, 1995). These biological nanomachines have captured the imagination of scientists and interested  
10   observers for over 300 years, since Antony Van Leeuwenhoek’s original description of “little legs”  
11   propelling the movement of microbes in 1675 (Dobell, 1958).

12           Flagella are structurally and functionally analogous to cilia and are most widely recognized as  
13   agents of cellular propulsion. Motile cilia and flagella drive the movement of many unicellular  
14   eukaryotes, including important human pathogens such as *Trichomonas vaginalis*, *Giardia lamblia* and  
15   *Trypanosoma brucei*, and are critical in the mating cycles of protozoa that cause malaria and  
16   toxoplasmosis (Ferguson, 2002; Vlachou et al., 2006). Cilia and flagella are also important for normal  
17   human physiology. For example, flagella drive sperm motility and motile cilia are critical for fluid  
18   movement across epithelial surfaces in the respiratory tract, brain ventricles, female reproductive tract  
19   and nodal cells of developing embryos (Ibanez-Tallon et al., 2003). Defects in these organelles lead to  
20   a wide variety of diseases and developmental defects (Badano et al., 2006). In addition to their obvious  
21   function in driving cell and fluid movement, flagella have recently been found to have an assortment of  
22   surprising functions beyond motility. For example, cilia and flagella are now recognized to play critical  
23   sensory and transport functions, and are important in cell cycle control. Several excellent reviews on

mammalian cilia and flagella are available (Badano et al., 2006; Satir and Christensen, 2007), and this review will focus on recent discoveries in the African trypanosome, *T. brucei*.

African trypanosomes are devastating human and animal pathogens. Two sub-species, *Trypanosoma brucei rhodesiense* and *T. b. gambiense* are causative agents of the fatal human disease known as African sleeping sickness. This disease is always fatal if left untreated, and it is estimated that 500,000 new infections occur annually (Legros et al., 2002; Welburn and Odiit, 2002). *T. brucei* is transmitted by the tsetse fly and alternates between bloodstream-form and insect-form life-cycle stages that are adapted to survive in the mammalian host and the insect vector, respectively. The importance of the flagellum for motility and for attachment to the tsetse fly salivary gland epithelium has been appreciated for many years (Vickerman, 1969, 1985; Vickerman and Preston, 1976). Indeed, the flagellum is responsible for the “undulating membrane” of bloodstream-form trypanosomes, which was one of the first described features of these parasites. Recent studies have revealed novel features of *T. brucei* flagellum structure, composition and function and these are outlined below. These discoveries are important not only from the standpoint of understanding trypanosome biology and the potential for uncovering novel drug targets, but also for advancing our understanding of fundamental aspects of eukaryotic flagellum structure and function.

## **2. Flagellum structure and composition**

The *T. brucei* cell body is roughly cylindrical in shape with tapered anterior and posterior ends. Culture-adapted insect midgut (procyclic) and bloodstream life-cycle stages used most commonly in the laboratory are approximately 19 - 25  $\mu$ m long (Robinson et al., 1995; Tyler et al., 2001), although the length of the cell body varies considerably during the life-cycle, particularly in the tsetse fly (Van Den Abbeele et al., 1999). A single flagellum emerges from the basal body near the posterior end of the cell. Once the flagellum exits the cytoplasm, it is surrounded by its own membrane and is attached along most of its length to the cell body, tracing a left-handed helical path from posterior to anterior

(Fig. 1). The distal end of the flagellum extends a short distance beyond the cell body. A specialized membrane domain termed the flagellar pocket forms from an invagination of the cell surface membrane where the proximal end of the flagellum emerges from the cytoplasm. The cell membrane, flagellar pocket membrane and flagellar membrane comprise three contiguous, but functionally distinct, membrane domains (Balber, 1990; Fridberg et al., 2007). Along the length of the cell, the flagellum and cell body are held in close apposition by a network of cytoskeletal and membranous connections that collectively make up the flagellum attachment zone (FAZ) (Vickerman, 1969; Kohl and Gull, 1998). Within the flagellum is a canonical “9 + 2” microtubule axoneme that drives flagellar movement. The flagellum also contains a large lattice-like structure, termed the paraflagellar rod (PFR), which runs parallel to the axoneme and is physically attached to both the axoneme and the FAZ.

### 2.1. Axoneme

As in most motile eukaryotic flagella, the trypanosome axoneme consists of nine outer doublet microtubules surrounding a pair of centrally located singlet microtubules. Radial spokes extend inward from each outer doublet toward the central pair. ATP-dependent dynein motor proteins attached to each doublet translocate along the adjacent doublet to generate the sliding forces that underlie flagellar movement. Connecting the outer doublets to one another are nexin links (Warner, 1976), which are thought to help maintain outer doublet organization (Lindemann and Kanous, 1997; Cibert, 2001) and may contribute to dynein regulation (Lindemann, 2004).

The ultrastructure of the trypanosome flagellum has been known for several decades, but only recently has the availability of genome sequences demonstrated that genes encoding axonemal proteins in other organisms are conserved in trypanosomes. This, combined with facile systems for RNA interference (RNAi) has spurred a burst in functional analyses of flagellar proteins in *T. brucei*. These studies have demonstrated conserved functions for several flagellum sub-complexes and provided

1 functional analysis of some proteins that were previously defined only by their biochemical association  
2 with the flagellum. These efforts have also uncovered novel and unexpected aspects of flagellum  
3 architecture in trypanosomes. Specifically, functional studies in *T. brucei* have revealed a requirement  
4 for  $\gamma$ -tubulin in the nucleation of central pair microtubules (McKean et al., 2003), uncovered a novel  
5 family of SMC-domain [“structural maintenance of chromosomes”] containing proteins as potential  
6 components of the nexin linkages (Fig. 2) (Baron et al., 2007b) and demonstrated a role for parkin co-  
7 regulated proteins in outer doublet microtubule stability (Fig. 3) (Dawe et al., 2005). RNAi directed  
8 against conserved components of the radial spokes, central pair apparatus (Fig. 4), outer dynein arms  
9 (Fig. 5), dynein regulatory complex and protofilament ribbons demonstrated a requirement for each of  
10 these structures in *T. brucei* flagellar motility (Ralston et al., 2005, 2006; Branche et al., 2006; Baron  
11 et al., 2007a). See Table 1 for a complete list of these proteins, as well as other characterized  
12 components of the *T. brucei* flagellum.

13         Analysis of mutants lacking central pair proteins also led to the surprising finding that the  
14 central pair retains a fixed orientation relative to outer doublet microtubules in *T. brucei* (Fig. 4)  
15 (Ralston et al., 2005, 2006; Branche et al., 2006). This differs from *Chlamydomonas reinhardtii*, a  
16 well-established model system for flagellum studies, where the central pair rotates within the axoneme  
17 (Omoto et al., 1999; Mitchell, 2003). Rotation of the asymmetrically arranged central pair apparatus in  
18 *C. reinhardtii* has been hypothesized to play a role in activating distinct sets of dyneins on the outer  
19 doublets (Wargo and Smith, 2003; Smith and Yang, 2004). Thus, a non-rotating central pair in *T.*  
20 *brucei* may reflect or impose unique demands on dynein motor regulation. Subsequent studies  
21 demonstrated that a fixed orientation of central pair microtubules is conserved among other  
22 kinetoplastids (Gadelha et al., 2006) and that defects in central pair orientation arise in some, but not  
23 all, flagellar mutants (Gadelha et al., 2006; Baron et al., 2007b). Thus central pair orientation appears  
24 to be actively maintained and is not simply the consequence of constraints imposed by flagellar beat in

trypanosomes. Interestingly, central pair orientation is also fixed relative to outer doublets in human axonemes, indicating that in some respects the trypanosome flagellum may provide advantages over *C. reinhardtii* as a model for human cilia.

## 2.2. Paraflagellar rod

One of the most peculiar structural features of the trypanosome flagellum is the presence of a large paracrystalline filament, the PFR, which extends alongside the axoneme from the flagellar pocket to the flagellum tip. Unlike the axoneme, which is broadly conserved among eukaryotes, the PFR is restricted to kinetoplastids, euglenoids and dinoflagellates. The PFR was first described 45 years ago (Vickerman, 1962b) and is required for normal motility (Bastin et al., 1998). However, its precise function is unknown and aside from its main structural components, PFR1 and PFR2 (Schlaeppli et al., 1989; Deflorin et al., 1994), and a few recently identified proteins (see below), its molecular composition remains to be determined.

Recently, several proteins affiliated with nucleotide metabolism have been found to be associated with the PFR. Three flagellar adenylate kinase proteins have been identified, with two of these (ADK-A and ADK-B) co-localizing with the PFR, and a third (ADK-E) localizing to the flagellum but not the PFR (Pullen et al., 2004; Ginger et al., 2005). Although adenylate kinase activity has not yet been demonstrated for these proteins, there appears to be adenylate kinase activity associated with the PFR since cytoskeletal extracts from the *snl-2* mutant, which lacks the majority of the PFR, have reduced adenylate kinase activity compared with wild-type extracts (Pullen et al., 2004). There is precedence for flagellar adenylate kinases as the central pair component *cpc1* of *C. reinhardtii* contains an adenylate kinase domain (Zhang and Mitchell, 2004). Two additional PFR-associated proteins involved in nucleotide metabolism, TbrPDEB1 and TbrPDEB2, were recently identified (Zoraghi and Seebeck, 2002; Oberholzer et al., 2007). These cAMP phosphodiesterases localize to the flagellum, though the majority of TbrPDEB2 localizes to punctuate spots in the cytoplasm (Oberholzer

et al., 2007). Knockdown of flagellar ADKs or PDEBs did not impair growth of procyclic cells, but simultaneous ablation of PDEB1 and 2 was found to be lethal in bloodstream-form parasites (Zoraghi and Seebeck, 2002; Ginger et al., 2005; Oberholzer et al., 2007). The motility phenotypes of PDEB knockdown mutants have not been described. Overall, these new findings suggest that the PFR is not simply a passive structural support to the axoneme, but might serve as a scaffold for assembly of regulatory and metabolic proteins that contribute to flagellum function.

### 2.3. *Flagellum attachment zone*

Components of the *T. brucei* FAZ are also mostly unknown, although the ultrastructure of the FAZ has been characterized in detail (Vickerman, 1962a; Sherwin and Gull, 1989; Gull, 1999; Kohl et al., 1999). The cytoplasmic side of the FAZ is defined by an electron-dense filament of unknown composition, together with four specialized microtubules that associate with a membranous compartment. A network of filaments connects the FAZ filament in the cell body to both the axoneme and PFR within the flagellum, and there are likely to be membranous linkages in addition to these cytoskeletal linkages between the cell body and flagellum (Hutchings et al., 2002). Flagellum adhesion glycoprotein 1 (Fla1), the *T. brucei* homolog of *T. cruzi* GP72, localizes to the FAZ and is required for flagellum attachment (el-Sayed et al., 1995; Nozaki et al., 1996; LaCount et al., 2000, 2002). Another FAZ component, FAZ1, was recently identified by screening an expression library with the monoclonal antibody L3B2 that recognizes the FAZ filament (Vaughan et al., 2008). FAZ1 is conserved amongst kinetoplastids and is required for normal FAZ assembly and flagellum attachment (Vaughan et al., 2008). Intriguingly, given the role of the FAZ in cytokinesis (see section 3.5), it is interesting that a *T. brucei* polo-like kinase homologue, TbPLK, has been localized to the FAZ (Kumar and Wang, 2006), although a separate study reported that TbPLK localizes to the cytoplasm (Hammarton et al., 2007).



#### 1    2.4. *Flagellar membrane and matrix*

2            The flagellar membrane and soluble flagellar matrix in *T. brucei* represent mostly uncharted  
3    territory. A single flagellum membrane-specific protein, the ESAG4 adenylate cyclase, has been  
4    identified in *T. brucei* (Paindavoine et al., 1992), and a calcium-binding flagellar membrane protein  
5    has been characterized in *T. cruzi* (Engman et al., 1989; Buchanan et al., 2005). In the flagellar matrix,  
6    several components of the intraflagellar transport pathway (IFT20, IFT52, IFT172) have recently been  
7    shown to exhibit both a soluble flagellar pool and a detergent-insoluble basal body-associated pool  
8    (Absalon et al., 2007a), but no other soluble flagellar proteins have been identified. Thus we have very  
9    little knowledge of the proteins that make up the matrix or membrane. For reference, proteomic  
10   analysis of the *C. reinhardtii* flagellum revealed 146 proteins with four or more unique peptides in the  
11   membrane + matrix fraction (Pazour et al., 2005). Given that the flagellar membrane presents an  
12   interface with the host (Webster and Russell, 1993; Fridberg et al., 2007), flagellar membrane proteins  
13   are likely to include critical components of signal transduction pathways that coordinate flagellar  
14   movement in response to environmental cues. Clearly, this represents an untapped area of research that  
15   is rich with opportunity for important discoveries.

16

#### 17   2.5. *Flagellum biogenesis*

18            Assembly of the *T. brucei* flagellum has been shown to be dependent on the conserved process  
19   of IFT (Rosenbaum, 2002; Kohl et al., 2003; Davidge et al., 2006). In this process, axoneme precursors  
20   are transported to the distal tip of the flagellum via IFT particles whose movement is powered by  
21   heterotrimeric kinesin (Kozminski et al., 1995). The DHC1b dynein then returns IFT particles to the  
22   base of the flagellum to reload with additional cargo (Pazour et al., 1999; Porter et al., 1999). Several  
23   conserved IFT components have now been identified and shown to function in *T. brucei* (Kohl et al.,  
24   2003; Davidge et al., 2006; Absalon et al., 2007a) and transport of an IFT52-GFP chimera has been  
25   directly visualized in live cells (Absalon et al., 2007a). Notably, since the trypanosome flagellum

exhibits many unique structural features, these parasites may also possess unique IFT machinery. Indeed, trypanosomes possess two DHC1b homologues, while other eukaryotes possess only a single retrograde IFT motor (Adhiambo et al., 2005). Likewise, putative IFT particles observed by electron microscopy in procyclic *T. brucei* cells are found almost exclusively adjacent to axonemal doublets 3, 4, 7 and 8 (Absalon et al., 2007a), a preference that has not been reported in other organisms.

In addition to the axoneme, flagellum biogenesis in *T. brucei* must accommodate assembly of the PFR. Construction of the PFR is dependent upon IFT (Kohl et al., 2003), although whether this reflects a direct involvement of IFT or a requirement for the axoneme is not clear. Consistent with the involvement of an IFT-like system for PFR assembly, PFR subunits are added primarily to the distal tip of the growing PFR filament (Bastin et al., 1999) and RNAi knockdown of the PFR2 subunit results in accumulation of the PFR1 subunit at the distal tip of the flagellum (Bastin et al., 1998).

The *T. brucei* flagellum persists throughout the cell cycle and the cell is faced with the challenge of elaborating a new daughter flagellum, while retaining the old one and co-ordinating all of this with other cell cycle events. Thus, flagellar biogenesis in these parasites poses unique challenges to the organism and is an exciting area that awaits further investigation. Interestingly, studies of flagellum assembly in *T. brucei* have led to the discovery of a novel quality control checkpoint, which functions to ensure proper assembly of tubulin dimers as they are delivered into the flagellum (Stephan et al., 2007).

## 2.6. Flagellum composition revealed through genomic and proteomic studies

In addition to the novel and surprising structural features uncovered by functional analysis of individual genes, significant advances have been made through large-scale genomic and proteomic approaches (Broadhead et al., 2006; Baron et al., 2007b). A recent comparative genomics study identified 50 genes that are conserved in *T. brucei* and other eukaryotes with motile flagella, but are absent in organisms that contain only non-motile flagella or lack flagella entirely. Hence these genes

1 represent core components of motile flagella (CMF) genes (Baron et al., 2007b). Of 50 *T. brucei* CMF  
2 genes identified, 30 were previously uncharacterized in any organism, providing an illustration of the  
3 number of surprises that await continued analysis of the trypanosome flagellum. RNAi knockdown of  
4 the 30 novel TbCMF genes, together with another 11 that had not previously been characterized in  
5 trypanosomes, demonstrated a motility function for the majority of these (Fig. 6). TbCMF proteins are  
6 expected to be axonemal, since they are conserved in organisms that lack a PFR or FAZ. Consistent  
7 with this, immunolocalization and biochemical fractionation demonstrated that several of these  
8 proteins are exclusively and stably associated with the flagellum, and ultrastructural analysis identified  
9 one family of novel TbCMF genes required for maintaining nexin linkages between axonemal outer  
10 doublets (Baron et al., 2007b).

11 The genomic analysis of Baron and colleagues also revealed that several flagellar genes are  
12 represented by families of two or more related sequences and that these gene families are often  
13 expanded in *T. brucei*. These trypanosome-specific gene expansions include components of the dynein  
14 regulatory complex, protofilament ribbons, dynein light chains and three novel protein families.  
15 Analysis of individual mutants and immunolocalization studies indicate that in at least some instances,  
16 this does not simply reflect functional redundancy (Baron et al., 2007b; K.L. Hill, unpublished  
17 observation). Trypanosomes also contain two homologues of the retrograde IFT molecular motor,  
18 DHC1b, which has a single orthologue in other eukaryotes. Trypanosome-specific expansion of  
19 flagellum gene families may reflect unique aspects of trypanosome flagellar beat, or may be a  
20 consequence of unique constraints imposed by the unusual architecture of the trypanosome flagellum.

21 Our inventory of trypanosome flagellar proteins was greatly expanded by proteomic analysis of  
22 salt-extracted flagella (Broadhead et al., 2006). This important study identified 331 proteins that  
23 constitute a *T. brucei*-extracted flagellum proteome (TbEFP). These proteins may be components of  
24 the axoneme, PFR, FAZ or basal body, which were all present in the sample used for analysis. Of the  
25 TbEFP dataset, 63% were kinetoplastid-specific and 26% were conserved only among flagellated

1 eukaryotes. RNAi knockdown indicated a flagellum function for eight TbEFP proteins while three  
2 others, including PFR2, had previously been shown to contribute to flagellum function (Bastin et al.,  
3 1998; Dawe et al., 2005). Interestingly, 238 of these proteins were annotated as hypothetical and there  
4 was a relatively small overlap (~50 proteins) between the trypanosome dataset and extracted axonemal  
5 proteomes from other organisms. This again emphasizes the unexplored nature of the trypanosome  
6 flagellar apparatus. These studies are also important from the standpoint of drug development, since  
7 trypanosome-specific flagellar proteins may represent novel targets for therapeutic intervention (see  
8 section 3.7).

9       An important outcome from functional genomic and proteomic studies in *T. brucei* was the  
10 identification of human disease gene candidates. Many human homologues of TbCMF and TbEFP  
11 genes have been directly implicated in human disease, or map to loci that are linked to diseases having  
12 known or suspected underpinnings of ciliary dysfunction (Broadhead et al., 2006; Baron et al., 2007b).  
13 For many of these genes, the works by Baron et al. and Broadhead et al. represent, to our knowledge,  
14 the first time that the connection to human disease has been made and, in some cases, the first  
15 functional characterization. For example, while it was known that Rib72 [protofilament ribbon protein  
16 72] was biochemically associated with flagellar protofilament ribbons (Patel-King, 2002; Ikeda and  
17 Kamiya, 2003), and that mutations in Rib72 are associated with juvenile myoclonic epilepsy (Suzuki et  
18 al., 2004; King, 2006), studies in *T. brucei* demonstrated a requirement for Rib72 in flagellar motility  
19 (Baron et al., 2007b). Similarly, mutations in hydin induce hydrocephalus and studies in *T. brucei* were  
20 among the first to show a specific flagellum function, demonstrating that hydin knockdown leads to  
21 defects in the positioning and formation/stability of the axonemal central pair (Broadhead et al., 2006;  
22 Dawe et al., 2007; Lechtreck and Witman, 2007). In another recent work, it was shown that two *T.*  
23 *brucei* homologues of a gene implicated in male sterility (Lorenzetti et al., 2004), PACRG, are  
24 required for outer doublet stability in *T. brucei* (Fig. 3) (Dawe et al., 2005). Thus, studies in *T. brucei*  
25 have added to the repertoire of genes implicated in human ciliary diseases and have provided

demonstration that these genes have bona fide flagellum functions. *T. brucei* is therefore emerging as an extremely powerful experimental system for probing candidate disease genes. It will become even more valuable as the era of gene discovery comes to a close and the focus turns to functional analysis.

### 3. Flagellum function

The *T. brucei* flagellum is a multifunctional organelle with many critical roles, some of which are well-established and some of which have only recently come to light. The flagellum has long been known for its role in driving parasite motility, which impacts both disease pathogenesis and transmission (Hill, 2003), and facilitating host cell attachment in the salivary glands of the tsetse fly vector (Vickerman, 1985; Vickerman et al., 1988). RNAi knockdown studies have now firmly established the central importance of the *T. brucei* flagellum and flagellar motility in other processes, including cell morphogenesis, cell division and immune evasion. Intriguingly, there is a dichotomy in the requirement for the flagellum during cell division between procyclic and bloodstream life-cycle stages, implying developmental regulation of flagellum function.

#### 3.1. Host cell attachment

Parasite attachment to host cells in the tsetse salivary gland is mediated by the flagellum and is a defining step in the developmental cycle of *T. brucei* and other salivarian trypanosomes. Although very little new information is available regarding host cell attachment, a brief discussion is provided to emphasize the need for more effort directed at understanding the underlying molecular and structural mechanisms. Parasite attachment to the salivary gland epithelium evokes extensive remodeling of the flagellar membrane and structural elements to establish intimate interdigitations between the parasite flagellum and tsetse salivary gland epithelial cells (Tetley and Vickerman, 1985; Vickerman, 1985; Tetley et al., 1987; Vickerman et al., 1988). These morphogenic events are associated with the final stages of parasite development into mammalian-infective trypomastigotes, including acquisition of the

variant surface glycoprotein (VSG) coat (Tetley and Vickerman, 1985; Vickerman, 1985; Tetley et al., 1987; Vickerman et al., 1988). Attachment to the salivary gland via the flagellum is therefore of obvious importance for parasite survival and pathogenesis. The molecular mechanisms and signaling events that underlie morphological changes and developmental transformation of the parasite upon host cell attachment are unknown and demand further attention.

### 3.2. Motility and flagellar beat

The *T. brucei* flagellum wraps around the cell body in a left-handed helix and flagellar beat causes the entire cell to rotate as it moves forward in an auger-like motion (Hill, 2003). The genus name *Trypanosoma* is in fact derived from the Greek words for auger, “trypanon”, and cell, “soma”. Until recently, we knew very little about the molecular mechanisms of flagellar beat in *T. brucei* and this was almost exclusively derived by extrapolation from other systems, with little or no direct experimental evidence from trypanosomes. While some aspects of axonemal motility can reasonably be expected to be conserved in *T. brucei*, other features are unique and warrant direct examination. For example, in addition to the structural peculiarities discussed above, the *T. brucei* flagellum moves rapidly in three dimensions, producing a spiral waveform with beat parameters that vary significantly along the length of the axoneme. Hence, flagellar beat in these parasites is highly dynamic and considerably more complex than in most other systems where it has been studied. Moreover, the dominant waveform in *T. brucei* and other trypanosomatids is a tractile beat that initiates at the tip of the flagellum and propagates toward the base (Walker, 1961; Walker and Walker, 1963), which is opposite of the base-to-tip beat observed in most other flagella. This tractile beat is disrupted intermittently with a brief base-to-tip beat, although this is not sustained in wild-type trypanosomes (Branche et al., 2006; Baron et al., 2007a).

Control of flagellar beat requires co-ordinate regulation of several thousand dynein motors having distinct subunit composition and functionality, which are arrayed along the inner and outer face

1 of axonemal doublet microtubules (Porter and Sale, 2000). RNAi knockdown of two separate dynein  
2 subunits have revealed that outer dynein motors are required for the tip-to-base beat that is a hallmark  
3 of *T. brucei* flagella (Branche et al., 2006; Baron et al., 2007a). Branche and colleagues (Branche et al.,  
4 2006) targeted the outer dynein intermediate chain DNAI1, while Baron et al. targeted the outer dynein  
5 light chain LC1 (Baron et al., 2007a). In both cases, loss of outer dynein motors (Fig. 5) resulted in  
6 loss of the tip-to-base beat, emergence of a sustained base-to-tip beat, and loss of forward motility.  
7 Reverse beat in LC1 mutants drove cell movement backward, with the flagellum tip trailing. These  
8 results provide clues into mechanisms controlling beat direction. They also illuminate an interesting  
9 contrast between trypanosomes and *Chlamydomonas*, where loss of outer arm dynein prevents flagella  
10 from assuming a symmetric base-to-tip beat (Kamiya and Okamoto, 1985). In other words, in one  
11 organism (trypanosomes), loss of the outer dyneins allows only symmetric base-to-tip beating, whereas  
12 in the other organism (*Chlamydomonas*), loss of outer dyneins prevents this beat form. Therefore, the  
13 data suggest that there are specialized functions of outer dynein motors in *T. brucei*.

14       There are also species-specific differences in the systems that regulate dynein activity. Ralston  
15 and co-workers (Ralston et al., 2005, 2006) recently showed that dynein regulation in *T. brucei* is  
16 mediated at least in part by a conserved dynein regulatory complex (DRC), which was originally  
17 discovered through genetic suppressor analysis in *C. reinhardtii* (Huang et al., 1982; Piperno et al.,  
18 1992; Gardner et al., 1994). The importance of the DRC is evidenced by the tumbling phenotype and  
19 loss of directional motility in DRC mutants (Hutchings et al., 2002; Rupp and Porter, 2003).  
20 Interestingly, although the DRC subunit trypanin is broadly conserved in flagellated eukaryotes (Hill et  
21 al., 2000; Ralston et al., 2005; Ralston and Hill, 2006), kinetoplastids are unique in having two  
22 trypanin-related proteins (Ralston et al., 2005, 2006). Recent data indicate that these proteins are not  
23 functionally redundant (K.L. Hill, unpublished observation), and it is reasonable to expect that  
24 constraints imposed by the unusual architecture or motility demands of the trypanosome flagellum may  
25 lead to unique requirements for dynein regulation in trypanosomes.

### 3.3. Endocytosis and immune evasion

The flagellar pocket is the sole site of endocytosis and secretion in *T. brucei* (Webster and Russell, 1993). The flagellar pocket is intimately associated with the flagellum and the external entrance to the flagellar pocket lumen is demarcated by close contacts between the flagellar and flagellar pocket membranes. Therefore, it has long been speculated that flagellar beating may influence trafficking through the flagellar pocket. Until recently however, experimental evidence in support of this idea has been lacking. Engstler and colleagues have now directly demonstrated that normal flagellar beat is necessary for uptake of surface Ig complexed with VSG in bloodstream-form *T. brucei* (Engstler et al., 2007). Since clearance of VSG-Ig protein complexes from the parasite surface is part of an integrated strategy for avoiding destruction by the host immune system, these studies suggest that flagellar motility is necessary for immune evasion by trypanosomes and establishment of a persistent infection in the mammalian host.

### 3.4. Cell morphogenesis

The trypanosome flagellum persists throughout the cell cycle and must be faithfully replicated and segregated along with other single copy organelles during cell division. Early indications that the flagellum itself might contribute to organelle positioning and cell morphogenesis came from structural studies showing close association between the basal body and the kinetoplast, suggesting that segregation of these single copy organelles might be obligately linked (Robertson, 1913; Simpson, 1968). Experimental evidence supporting this idea came from the demonstration that the basal body and kinetoplast are indeed physically interconnected and that pharmacological perturbation of basal body segregation blocks kinetoplast segregation (Robinson and Gull, 1991). These studies provided an important framework for considering the role of the cytoskeleton in mitochondrial genome segregation (Robinson and Gull, 1991; Robinson et al., 1995), though they did not directly implicate the flagellum



1 as an active player in this process. Direct evidence for flagellum growth and flagellar beat in  
2 kinetoplast segregation has now come from RNAi studies in which several independent mutants with  
3 defective flagellum biogenesis or movement have a common defect in kinetoplast positioning (Dawe et  
4 al., 2005; Ralston et al., 2005, 2006; Branche et al., 2006; Absalon et al., 2007b). At present it is still  
5 unclear precisely how the flagellum effects kinetoplast segregation, but a model (Absalon et al.,  
6 2007b) has been proposed in which the elongating new flagellum, which is tethered at the growing end  
7 by the flagellar connector (Moreira-Leite et al., 2001), exerts a reaction force on the basal body,  
8 driving movement of the new basal body/kinetoplast away from the old one (Absalon et al., 2007b).  
9 Given the central importance of the flagellum and flagellar motility to trypanosome biology, it seems  
10 advantageous that segregation of the mitochondrial genome is linked to flagellum formation,  
11 maintenance and function.

12       The discovery of a novel mobile connector between the old and new flagellum, termed the  
13 flagellar connector(FC) (Moreira-Leite et al., 2001), suggests that flagellum influence on positioning  
14 of organelles extends beyond the kinetoplast. The flagellar connector is a mobile, tripartite cytoskeletal  
15 structure that connects the tip of the new flagellum to the old flagellum as it grows in parallel to this  
16 old flagellum. Hence, the old flagellum defines the placement of the new flagellum and by extension is  
17 suspected to impart positional information that ensures correct polarity and organization of the  
18 daughter cell. The FC has been visualized by electron microscopy (Moreira-Leite et al., 2001) and  
19 antibody staining (Briggs et al., 2004), however neither the protein components of the FC nor the  
20 mechanism for FC movement are currently known. Interestingly, the FC appears to be specific to the  
21 procyclic life-cycle stage of *T. brucei*, since it has not been observed in the bloodstream-form stage or  
22 in other trypanosomatids (Briggs et al., 2004).

### 24 3.5. Cell division

1 African trypanosomes divide through binary fission. The cleavage furrow forms at the anterior  
2 end of the cell, between the new and old flagellum. The furrow then advances toward the cell posterior,  
3 dividing the cell into two daughter cells that are oriented with their flagella facing in opposite  
4 directions just before final cell separation occurs (Fig. 7) (Sherwin and Gull, 1989). It has previously  
5 been suggested that the flagellum and FAZ provide positional and directional cues for cleavage furrow  
6 formation (Robinson et al., 1995). RNAi knockdown of genes required for flagellum biogenesis,  
7 structure and movement have now provided direct evidence in support of this hypothesis. In a seminal  
8 study by Bastin and colleagues (Kohl et al., 2003), RNAi knockdown of IFT genes in procyclic cells  
9 not only showed that IFT is required for flagellum biogenesis in *T. brucei*, but also demonstrated that  
10 the flagellum is an essential organelle in these parasites. IFT knockdown halts new flagellum growth,  
11 resulting in cells with a new flagellum and FAZ that are shorter than normal. These cells initiate  
12 cytokinesis at a point that correlates with the position of the truncated FAZ filament, giving rise to  
13 progressively shorter cells and ultimately yielding aflagellate cells that are non-viable. Cells  
14 completely lacking flagella have disorganized organelles and abnormal cell shape. Therefore, in  
15 addition to influencing the position of intracellular organelles, the flagellum and FAZ define the  
16 position and direction of cleavage furrow ingression during cytokinesis (Fig. 8). Consistent with the  
17 importance of the FAZ in cytokinesis, RNAi knockdown of the FAZ proteins Fla-1 and FAZ1 lead to  
18 flagellum detachment and cytokinesis failure (LaCount et al., 2002; Vaughan et al., 2008).

19 The potential to control cleavage furrow formation and cell size based on FAZ positioning has  
20 direct relevance to trypanosome development, since large variations in cell size occur as part of the  
21 natural life-cycle of *T. brucei* (Van Den Abbeele et al., 1999). For example, transformation from  
22 trypomastigotes to epimastigotes during development in the tsetse fly vector is associated with an  
23 assymetric cell division that yields one long and one short daughter cell (Van Den Abbeele et al.,  
24 1999). Recent studies have now shown that reduced cell size accompanying this differentiation is  
25 correlated with flagellum length and position in the newly forming daughter cell (Sharma et al., 2008).

1 Since this asymmetric division occurs only after the parasite reaches specific host tissues, control of  
2 cell cleavage by the FAZ/flagellum is likely regulated in response to specific environmental cues.

### 3 4 3.6. *Flagellar motility and cytokinesis*

5 Perhaps even more surprising than the role for flagellum assembly and attachment in cleavage  
6 furrow formation was the finding that flagellar beating is required to complete cell division in  
7 procyclic cells (Fig. 9) (Ralston et al., 2005, 2006; Branche et al., 2006). RNAi knockdown of radial  
8 spoke and central pair proteins in the procyclic life-cycle stage leads to cells with full-length, attached  
9 flagella that are almost completely immotile (Ralston et al., 2005, 2006; Branche et al., 2006). These  
10 mutants fail in the final stage of cytokinesis, giving rise to daughter cells that remain connected at their  
11 extreme posterior ends (Ralston et al., 2005, 2006; Branche et al., 2006). Continued rounds of  
12 cytokinesis initiation in the absence of cell separation results in large, multicellular clusters. This cell  
13 division defect has now been observed in 33 independent motility mutants in the procyclic life-cycle  
14 stage (Ralston et al., 2005, 2006; Branche et al., 2006; Baron et al., 2007b) and the severity of the cell  
15 separation defect correlates with the severity of the motility defect (Baron et al., 2007b). Moreover, the  
16 block in cell division can be rescued by mechanical agitation (Ralston et al., 2005, 2006; Branche et  
17 al., 2006). The combined data therefore indicate that physical forces supplied by flagellar beating  
18 contribute to normal cytokinesis in procyclic cells. In wild-type *T. brucei*, newly replicated daughter  
19 cells are oriented with their flagella pointing in opposite directions and eventually pull apart (Fig. 7).  
20 Hence, the pulling and rotational forces provided by flagellar beating may provide a convenient way to  
21 drive the final steps of cell separation which, in trypanosomes, occurs in the absence of any detectable  
22 contractile ring. A similar phenomenon, termed rotokinesis, has been observed in *Tetrahymena* (Brown  
23 et al., 1999). Therefore, in addition to its role as a spatial and directional cue for directing cleavage  
24 furrow formation, the flagellum plays an active role in cell division in procyclic *T. brucei* cells.

### 3.7. Flagellar motility and cytokinesis in bloodstream-form cells

The vast majority of work on flagellum structure and function in *T. brucei* has been carried out in procyclic cells. Three important studies have now demonstrated that the requirement of the flagellar apparatus in cytokinesis extends to bloodstream-form cells (Fig. 9) (Branche et al., 2006; Broadhead et al., 2006; Ralston and Hill, 2006). Two studies used inducible RNAi to block expression of flagellar proteins in bloodstream-form cells. Ralston and Hill (2006) targeted the DRC subunit trypanin, while Broadhead and colleagues (Broadhead et al., 2006) targeted PFR2, MBO2, PACRG-A and two novel proteins within the TbEFP dataset. In a third study, Branche and colleagues targeted PFR2 for transient knockdown by transfection of double-stranded RNA (dsRNA) (Branche et al., 2006). In all cases these flagellar proteins were found to be essential. This was a surprising result, since none of these proteins are essential in the procyclic life cycle stage (Bastin et al., 1998; Hutchings et al., 2002; Broadhead et al., 2006). The terminal phenotype in all mutants was striking (Fig. 9b). Mutants failed to initiate cytokinesis but continued through multiple rounds of organelle replication, accumulating as large, amorphous masses with multiple kinetoplasts, flagella and nuclei. In at least one case, the kinetics of phenotype progression was shown to correlate with the extent of knockdown (Ralston and Hill, 2006). In procyclic *T. brucei* cytokinesis defects can arise from abnormal formation and/or positioning of the flagellum, FAZ or kinetoplast/basal body complex (see sections 3.4 and 3.5). However, in the work by Ralston, each of these events was examined and found to occur as expected (Fig. 9c) (Ralston and Hill, 2006). Importantly, Broadhead and colleagues showed that bloodstream-form flagellar mutants were endocytically active (Broadhead et al., 2006). Therefore, the cytokinesis defect and lethality in bloodstream-form flagellum mutants is not due to failed organelle replication or positioning or a block in endocytosis. Rather, the phenotype is a direct consequence of perturbing flagellum structure and/or function.

The above studies demonstrate that the requirement for flagellar apparatus in cytokinesis is a common feature of procyclic and bloodstream-form *T. brucei*. Nonetheless, there are important

1 differences between procyclic and bloodstream-form flagellum mutants that must be considered.  
2 Firstly, bloodstream-form cells are more sensitive to flagellum defects, since none of the proteins  
3 examined in bloodstream-form cells are essential in procyclic cells (Fig. 9b). Moreover, PACRG-A  
4 knockdown produces no discernable phenotype in procyclic cells (Dawe et al., 2005), but is lethal in  
5 the bloodstream-form (Broadhead et al., 2006). In the case of trypanin, procyclic mutants are viable  
6 with little or no effect on cell doubling even though trypanin is undetectable (Hutchings et al., 2002),  
7 while bloodstream-form cells die when protein levels are reduced two to four-fold. Second, mechanical  
8 agitation rescues the cytokinesis defect of procyclic mutants (Ralston et al., 2005, 2006; Branche et al.,  
9 2006) but not of bloodstream-form mutants (Griffiths et al., 2007; K.L. Hill, unpublished observation).  
10 Third, the terminal phenotype of flagellar mutants is different between life cycle stages (Fig. 9a and  
11 9b). Procyclic mutants fail during the progression and/or completion of cell cleavage. Bloodstream-  
12 form mutants ultimately fail to initiate cytokinesis. These results suggest life-cycle stage-specific  
13 regulation of flagellum function and emphasize the emerging concept (Hammarton et al., 2003; Tu and  
14 Wang, 2004) that there is a dichotomy with respect to factors that control cell division in bloodstream-  
15 form and insect-form African trypanosomes.

16       The combined data indicate that the flagellum and perhaps flagellar motility itself is essential in  
17 bloodstream-form *T. brucei*. Note however, that while these studies show that flagellar proteins are  
18 essential, they do not demonstrate that motility itself is essential. For example, it is possible that  
19 lethality in bloodstream flagellar mutants results from a structural defect, rather than a defect in  
20 motility, per se. This is more than a remote possibility for several reasons. Firstly, procyclic trypanin  
21 mutants have an actively beating flagellum (Hutchings et al., 2002) and PACRG-A procyclic mutants  
22 were reported to show no discernable motility defect (Dawe et al., 2005). Yet, both of these mutants  
23 were inviable in the bloodstream stage. Therefore, lethality in bloodstream-form mutants does not  
24 strictly correlate with motility defects in the corresponding procyclic mutants. Second, most of the  
25 mutants examined are known to produce structural changes in flagellum sub-structures, as directly

1 shown in *T. brucei*, or in the analogous *C. reinhardtii* mutants. For example, PFR2 mutants disrupt the  
2 PFR, trypanin mutants disrupt the DRC, and MBO2 mutants loose the beak structure within the lumen  
3 outer doublet microtubules. Two other proteins targeted in bloodstream-form cells, LC1 and Tax1, are  
4 dynein subunits (Yamamoto et al., 2006; Baron et al., 2007b). Work in *T. brucei* (Branche et al., 2006;  
5 Baron et al., 2007a) and *C. reinhardtii* (Sakato and King, 2004) demonstrate that loss of dynein  
6 subunits causes disruption of the entire axonemal dynein arm. Since detailed ultrastructural analysis  
7 has not been completed on any of the bloodstream-form mutants, the structural consequences of  
8 knockdown in this life cycle stage is unknown. Finally, since there is an emerging link between  
9 flagellum assembly/disassembly and cell cycle control in other organisms, it is possible that perturbing  
10 fidelity of the flagellum, rather than motility itself, triggers a cytokinesis checkpoint. Therefore, it  
11 remains to be determined whether or not flagellar motility is essential in bloodstream-form  
12 trypanosomes. Clearly, many more exciting discoveries can be expected from continued investigation  
13 of the connection between the flagellar apparatus and cell division in these deadly pathogens.

14       Regardless of the precise reason for lethality in bloodstream-form flagellum mutants, the above  
15 studies raise the very exciting possibility that the flagellum might be exploited as a novel target for  
16 therapeutic intervention in African sleeping sickness. The flagellum is comprised of an estimated 500 –  
17 600 proteins (Pazour et al., 2005; Merchant et al., 2007), including many with enzymatic activity  
18 and/or small ligand binding sites. Such small molecule-binding proteins make up a large fraction of  
19 proteins considered to be good drug targets (Hopkins and Groom, 2002). Therefore, an essential role  
20 for flagellar proteins in bloodstream-form *T. brucei* greatly enhances the potential of this organelle as a  
21 novel drug target. In order to exploit this potential, it will be critical to determine the precise structural  
22 and enzymatic features of flagellar proteins and flagellar sub-complexes in these parasites.

23

#### 24 **4. Summary and future considerations**

1       The flagellum of African trypanosomes is now established as an essential and multifunctional  
2 organelle with critical roles in host cell attachment, cell motility cell morphogenesis, cell division and  
3 immune evasion. Moreover, all flagellum structural proteins so far examined have been found to be  
4 essential in the bloodstream life-cycle stage, indicating that the flagellum may represent a rich source  
5 of novel targets for therapeutic intervention in African sleeping sickness. Much work is still needed to  
6 capitalize on this possibility and to determine the exact nature of the requirement for the flagellum and  
7 flagellar proteins in trypanosome cell division and cell polarity determination. Further surprises almost  
8 certainly await continued investigation, particularly in the area of flagellar membrane and matrix  
9 composition, IFT and flagellar biogenesis. It is therefore likely that this old dog may still have a few  
10 new tricks to teach us.

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## Figure legends

Fig. 1. The trypanosome flagellum. (A) Scanning electron micrograph of a procyclic trypanosome. An arrow indicates the single flagellum that is attached along the cell body. (B) Transmission electron micrograph, showing a cross-section of the flagellum and its attachment to the cell body as viewed from posterior toward anterior. (C) Schematic representation of the micrograph in B. Major flagellar sub-structures are indicated and outer doublet microtubules are numbered according to conventional nomenclature. The four specialized sub-pellicular microtubules that are part of the FAZ are indicated in blue. Abbreviations: DRC, dynein regulatory complex; IFT, intraflagellar transport; FAZ, flagellum attachment zone; PFR, paraflagellar rod.

Fig. 2. RNA interference (RNAi) knockdown of the CMF9 (conserved component of motile flagella 9) protein results in loss of inter-doublet connections. Transverse (A, B) and longitudinal (C) sections of procyclic cells in which CMF9 is knocked down with RNAi. White arrows point to misplaced outer doublets. Scale bars are 100 nm (A), 50 nm (B) and 200 nm (C). Adapted from Baron et al., 2007b, with permission.

Fig. 3. Simultaneous RNA interference (RNAi) knockdown of the parkin co-regulated genes PACRGA and PACRGB results in loss of outer doublets. Serial transverse sections (proceeding from posterior to anterior (A, B, C, respectively) of a procyclic cell in which both PACRGA and B have been knocked down with RNAi. Black arrows indicate the outer doublet that is lost toward the distal tip of the axoneme. The white arrow indicates the electron dense lumen of the A-tubule. Adapted from Dawe et al., 2005, with permission.

Fig. 4. Central pair orientation is fixed in wild type cells and is dependent upon the central pair protein PF16 [paralyzed flagella 16]. (A, B) Transverse sections of PF16 knockdowns grown in the absence (-) or presence (+) of tetracycline to induce RNAi knockdown of PF16. In control cells (- Tet) central pair microtubules lie in a plane that is roughly parallel to the paraflagellar rod, whereas in PF16 knockdown mutants (+ Tet) central pair orientation is variable. (C, D) Schematic with lines depicting the plane of central pair microtubules measured in control (C) and PF16 knockdown (D) cells,  $n = 26$ . Outer doublet number 1 is labeled for reference. Scale bar is 100 nm. Adapted from Ralston et al., 2006, with permission. [This figure will be printed in black and white for hard copy. –will the online version be in color?]

Fig. 5. RNA interference (RNAi) knockdown of LC1 (dynein light chain 1) or DNAI1 (axonemal dynein intermediate chain 1) results in loss of outer dynein arms. Transverse sections of flagella (A, B) or flagellar cytoskeleton (C) from LC1 (A, B) or DNAI1 (C) knockdown lines grown in the absence (-) or presence (+) of tetracycline. Arrows point to positions of outer dynein arms, which are absent in LC1 and DNAI1 knockdowns. Scale bar is 100 nm. Adapted from Baron et al., 2007a (A) and Branche et al., 2006 (C), with permission. [Do you think black arrows might be clearer in this figure?]

Fig. 6. Importance of flagellum motility revealed by procyclic motility mutants. Schematic representations of motility phenotypes in procyclic *T. brucei* flagellum mutants. Wild-type (WT) cell movement is helical with the flagellum tip leading. RNA interference knockdown of flagellar protein expression leads to a variety of motility phenotypes. “Dizzy” mutants, such as trypanin knockdown mutants (Hutchings et al., 2002), retain a vigorously beating flagellum, but are unable to move directionally, and instead tumble in place. “Reverse” motility is observed when subunits of the outer dynein motors, such as DNAI1 (Branche et al., 2006) or LC1 (Baron et al., 2007a) are targeted. These mutants exhibit a reverse, base-to-tip beat and move backward with the flagellum tip trailing.

1 “Paralyzed” mutants, such as central pair and radial spoke mutants (Branche et al., 2006; Ralston et al.,  
2 2006) are incapable of movement at the cellular level. Some paralyzed mutants have flagella that are  
3 able to beat, while other mutants have more severe defects and are only capable of erratic twitching  
4 motions. “Lethal” defects arise in mutants with severe flagellar paralysis (Baron et al., 2007b).

5  
6 Fig. 7. Cartoon depicting major stages of the *T. brucei* cell division cycle in procyclic lifecycle stages.  
7 The cell cycle begins with replication of the basal body and initiation of new flagellum (blue)  
8 biogenesis. The new flagellum extends along a path defined by the old flagellum (green), while the  
9 basal bodies and associated kinetoplasts (small gray circles) migrate away from each other. In a closed  
10 mitotic cycle, the nucleus (large gray circle) is replicated and the new nucleus assumes a position  
11 between the new and old kinetoplast/basal body apparatus. Cytokinesis initiates at the tip of the new  
12 flagellum (black arrowhead) and cleavage furrow ingression proceeds to the cell posterior, separating  
13 the two daughter cells between the new and old flagellum. Ultimately, daughter cells are oriented in  
14 opposite directions, with their flagella exerting rotational and pulling forces that facilitate final cell  
15 separation.

16  
17 Fig. 8. Flagellum attachment zone (FAZ) length defines cell cleavage site. (A, B)  
18 Immunofluorescence (A) and merged (B) image of a dividing procyclic cell following RNA  
19 interference knockdown of IFT88 [intraflagellar transport protein 88], showing the paraflagellar rod  
20 (green), FAZ (red) and nucleus (blue). (C) Cartoon depicting the result of improper cell cleavage in an  
21 IFT88 knockdown (left), resulting in one normal-sized daughter cell (right, top) and one short daughter  
22 cell (right, bottom). The site of cell cleavage corresponds to the end of the new FAZ filament (white  
23 arrowheads in A and B. Adapted from Kohl et al., 2003, with permission.

1 Fig. 9. There is a dichotomy in the requirement of the flagellum in procyclic versus bloodstream form  
2 cells. (A) DIC images of uninduced (-Tet) and tetracycline-induced (+Tet) procyclic *pfr2* knockdown  
3 mutants. Induced cells fail in the completion of cytokinesis, and accumulate as clusters of distinct cell  
4 bodies that remain physically attached at their posterior ends (arrowhead). (B) Phase-contrast images  
5 of uninduced and tetracycline-induced trypanin knockdown mutants in the procyclic (PCF) and  
6 bloodstream (BSF) lifecycle stages. Procyclic trypanin mutants are viable with little or no division  
7 defect, while bloodstream mutants fail in the initiation of cytokinesis and accumulate as amorphous  
8 masses with multiple flagella. (C) Immunofluorescence images of uninduced and tetracycline-induced  
9 bloodstream form trypanin mutants. Phase contrast images are shown in the left panels and staining for  
10 the FAZ (red) and DNA (blue) is shown in the right panels. Uninduced cells undergoing division have  
11 a maximum of two FAZ structures, which have a parallel configuration until cytokinesis is initiated.  
12 Induced cells have more than two parallel FAZ structures, which retain a parallel configuration,  
13 indicating that failure to initiate cytokinesis in these cells is not due to failed organelle replication or  
14 positioning. Adapted from Ralston et al., 2006 and Ralston and Hill, 2006, with permission.

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16

1 Table 1. Characterized components of the *Trypanosoma brucei* flagellum. This table lists  
2 proteins for which experimental investigation has allowed protein assignment to specific  
3 flagellar functions or flagellar sub-complexes in *T. brucei*.

Protein	Function and/or Location	Reference
<b>Axoneme</b>		
CMF9	OD Assembly/Maintenance	(Baron et al., 2007b)
CMF76b	OD Assembly/Maintenance	(Baron et al., 2007b)
Hydin	CP Maintenance/Orientation	(Dawe et al., 2007)
MBO2	OD Component	(Broadhead et al., 2006)
PACRG-A	OD Assembly/Maintenance	(Dawe et al., 2005)
PACRG-B	OD Assembly/Maintenance	(Dawe et al., 2005)
PF16	CP Orientation	(Branche et al., 2006; Ralston et al., 2006)
PF20	CP Orientation	(Branche et al., 2006; Ralston et al., 2006)
Rib72	OD Component	(Baron et al., 2007b)
RSP3	RS Assembly/Maintenance	(Ralston et al., 2006)
Trypanin	DRC Component, Dynein Regulation	(Ralston et al., 2006)
$\gamma$ -Tubulin	CP Assembly/Maintenance	(McKean et al., 2003)
<b>Dyneins</b>		
DHC1b-1	IFT	(Kohl et al., 2003; Adhiambo et al., 2005)
DHC1b-2	IFT	(Kohl et al., 2003; Adhiambo et al., 2005)
DNAI1	ODA	(Branche et al., 2006)
LC1	ODA	(Baron et al., 2007a)
Tax-1/p38	IDA	(Broadhead et al., 2006; Yamamoto et al., 2006)
<b>Paraflagellar Rod</b>		
ADK-A	Adenylate Kinase	(Pullen et al., 2004; Ginger et al., 2005)
ADK-B	Adenylate Kinase	(Pullen et al., 2004; Ginger et al., 2005)
PDEB1	cAMP Homeostasis	(Oberholzer et al., 2007)
PDEB2	cAMP Homeostasis	(Oberholzer et al., 2007)
PFR1	PFR Structural Protein	(Deflorin et al., 1994; Durand-Dubief et al., 2003)
PFR2	PFR Structural Protein	(Schlaeppli et al., 1989; Bastin et al., 1998)
Tryp-CaM	Ca <sup>2+</sup> binding	(Ridgley et al., 2000)
5.20	PFR component	(Woodward et al., 1994)
<b>Flagellum Attachment Zone</b>		
FAZ1	Flagellum Attachment	(Vaughan et al., 2008)
Fla1	Flagellum Attachment	(LaCount et al., 2002)
<b>Flagellar Membrane</b>		
ESAG4	Adenylate Cyclase	(Paindavoine et al., 1992)
<b>Intraflagellar Transport</b>		
Che2/IFT80	IFT	(Davidge et al., 2006; Absalon et al., 2007a)
IFT20	IFT, Flagellar Matrix	(Absalon et al., 2007a)
IFT52	IFT, Flagellar Matrix	(Absalon et al., 2007a)
IFT57/55	IFT	(Absalon et al., 2007a)
IFT88	IFT	(Kohl et al., 2003; Absalon et al., 2007a)

IFT122	IFT	(Absalon et al., 2007a)
IFT140	IFT	(Absalon et al., 2007a)
IFT172	IFT, Flagellar Matrix	(Absalon et al., 2007a)
PIFTA1	IFT	(Absalon et al., 2007a)
PIFTB2	IFT	(Absalon et al., 2007a)
PIFTC3	IFT	(Absalon et al., 2007a)
PIFTD4	IFT	(Absalon et al., 2007a)
PIFTF6	IFT	(Absalon et al., 2007a)

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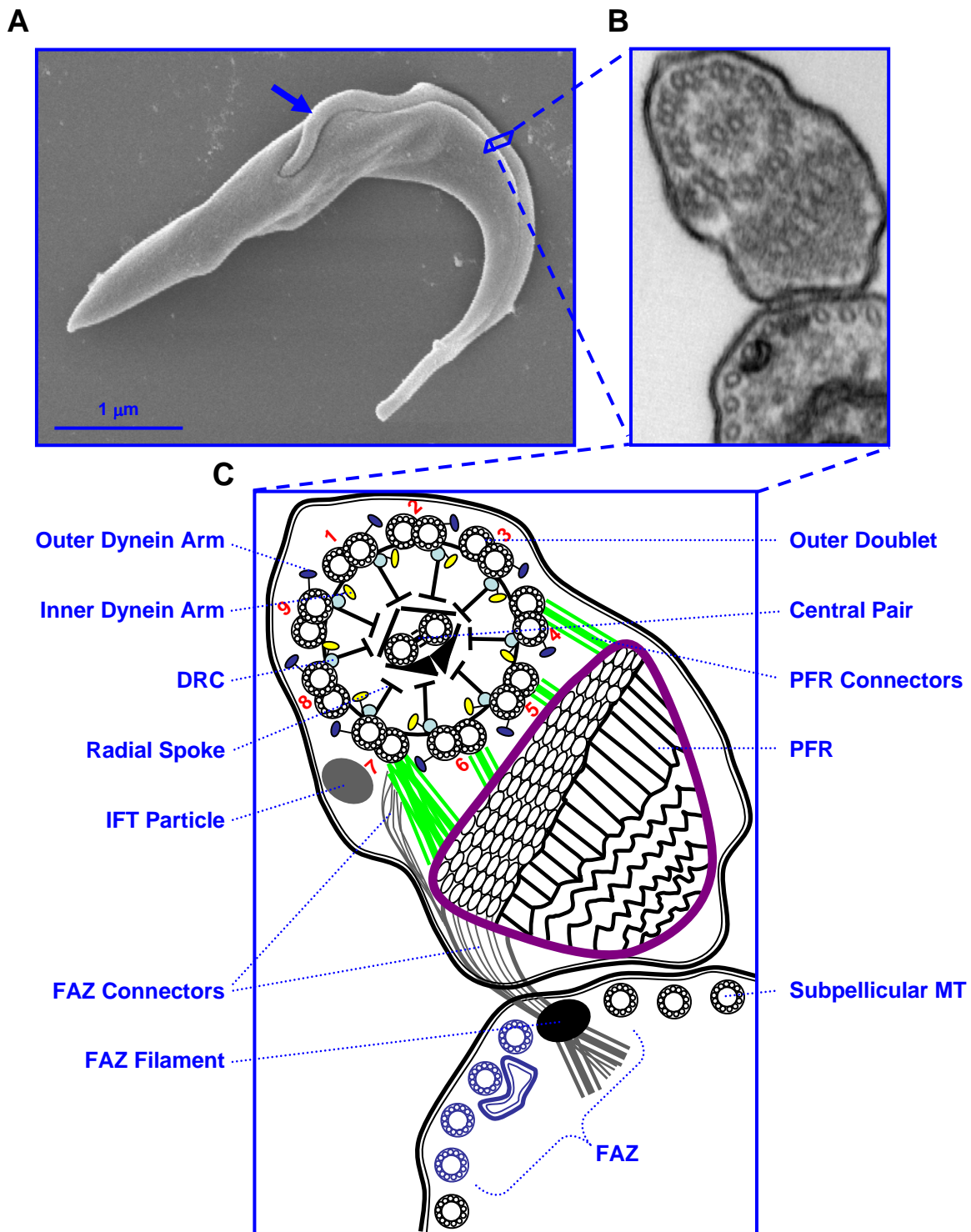
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2 While many additional novel flagellar proteins have been identified through comparative  
3 genomics (Baron et al., 2007b) and proteomics (Broadhead et al., 2006), and have been  
4 linked to flagellar functions by mutant or RNAi analyses, their precise functions and locations  
5 of are unknown. For a comprehensive inventory of flagellar proteins in a variety of organisms,  
6 the reader is referred to the ciliome database: [www.ciliome.com](http://www.ciliome.com) (Inglis et al., 2006).

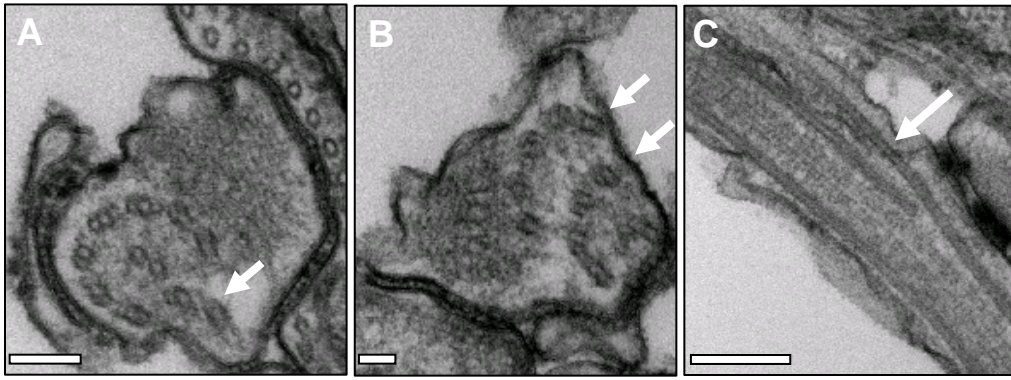
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8 Abbreviations: OD: outer doublet; CP: central pair; RS: radial spokes; IFT: intraflagellar  
9 transport; ODA: outer dynein assembly; IDA: inner dynein assembly; PFR: paraflagellar rod;  
10 DRC: dynein regulatory complex.

**Figure 1**

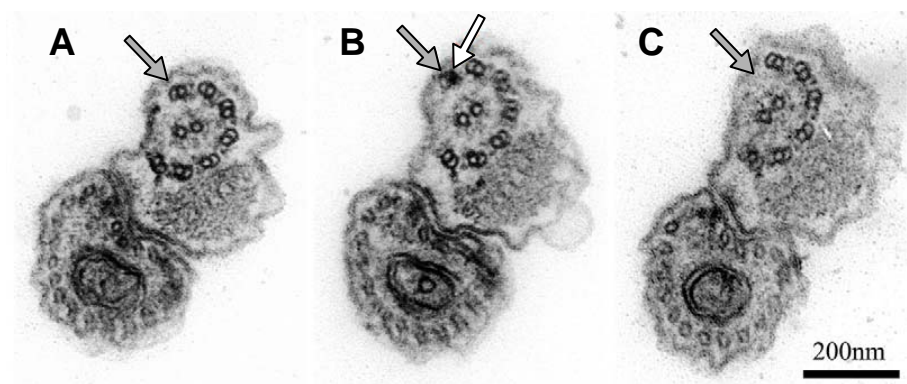


**Figure 2**

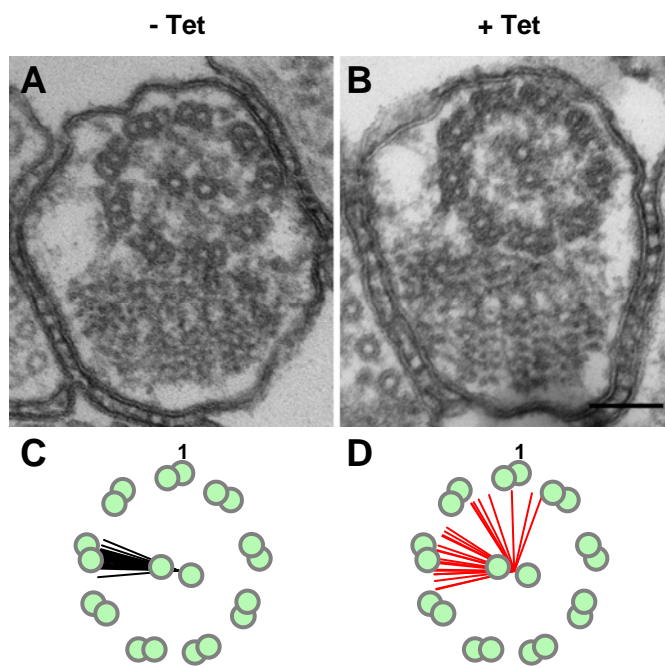




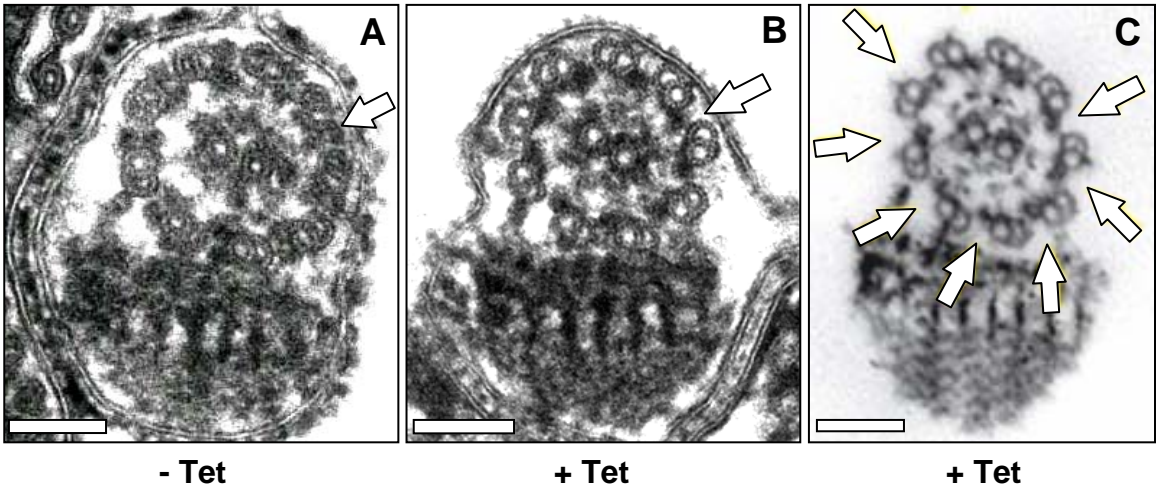
**Figure 3**



**Figure 4**



**Figure 5**



**Figure 6**

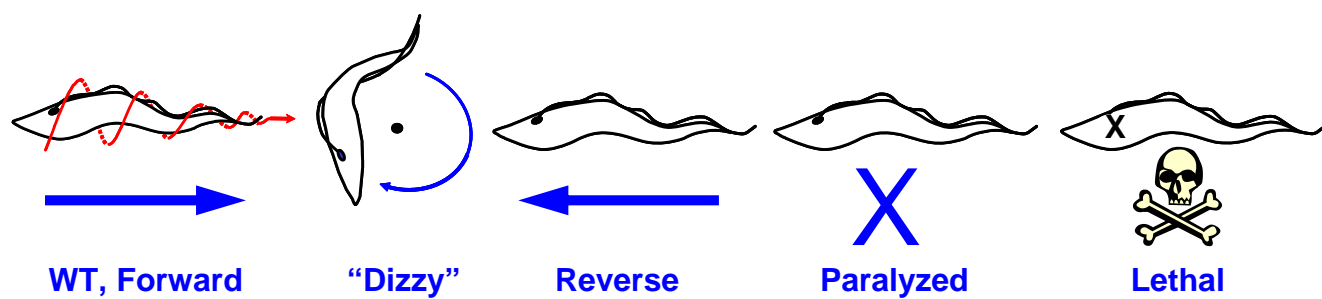


Figure 7

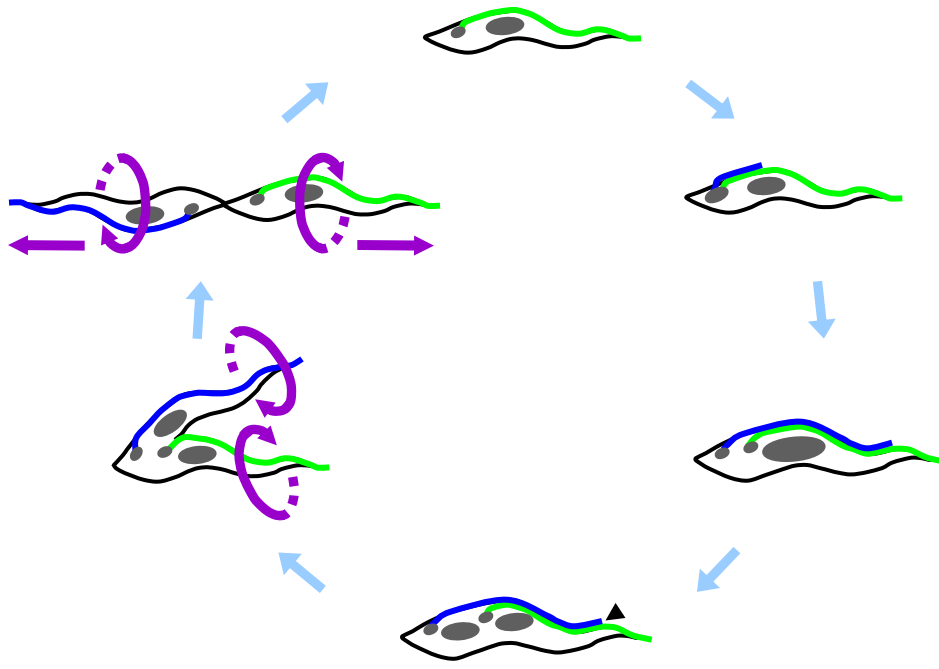
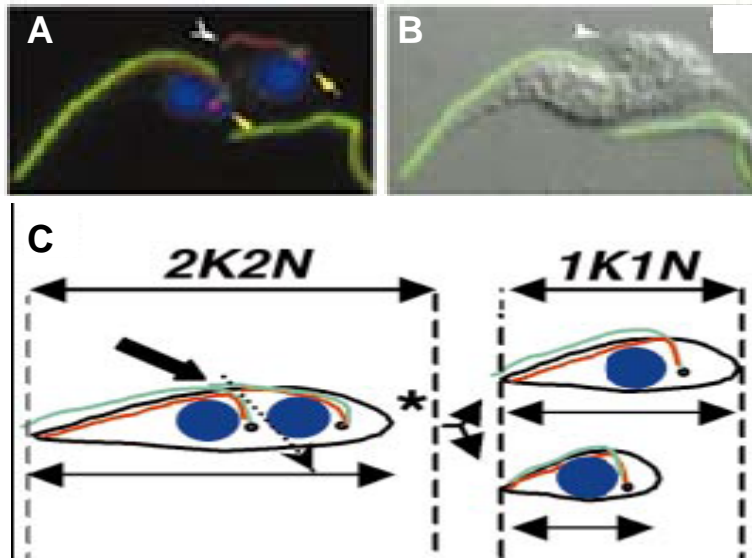


Figure 8



**Figure 9**

