

# Environmental influences in the evolution of tetrapod hearing sensitivity and middle ear tuning

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**Synopsis** Vertebrates inhabit and communicate acoustically in most natural environments. We review the influence of environmental factors on the hearing sensitivity of terrestrial vertebrates, and on the anatomy and mechanics of the middle ears. Evidence suggests that both biotic and abiotic environmental factors affect the evolution of bandwidth and frequency of peak sensitivity of the hearing spectrum. Relevant abiotic factors include medium type, temperature, and noise produced by nonliving sources. Biotic factors include heterospecific, conspecific, or self-produced sounds that animals are selected to recognize, and acoustic interference by sounds that other animals generate. Within each class of tetrapods, the size of the middle ear structures correlates directly to body size and inversely to frequency of peak sensitivity. Adaptation to the underwater medium in cetaceans involved reorganization of the middle ear for novel acoustic pathways, whereas adaptation to subterranean life in several mammals resulted in hypertrophy of the middle ear ossicles to enhance their inertial mass for detection of seismic vibrations. The comparative approach has revealed a number of generalities about the effect of environmental factors on hearing performance and middle ear structure across species. The current taxonomic sampling of the major tetrapod groups is still highly unbalanced and incomplete. Future expansion of the comparative evidence should continue to reveal general patterns and novel mechanisms.

## Environmental factors and hearing

In most tetrapods, the sense of hearing plays a fundamental role in predator avoidance, mate acquisition, and foraging (Dusenbery 1992). In support of such behaviors, hearing systems have adapted to a wide diversity of life histories such that the ears operate under a myriad of combinations of environmental conditions, sensitivity requirements, and size (Webster et al. 1992; Fay and Popper 1994, 1998; Dooling et al. 2000). Comparative analyses of this diversity can reveal correlations between morphological or mechanistic features of the ear and environmental factors in the habitat, indicating potential cases of adaptation.

In this review, we focus on the frequency range of hearing in tetrapods and examine the current evidence for adaptation to environmental factors. We also review the knowledge on changes to middle ear anatomy in response to environment. While inner ear allometry, mechanics, and neurophysiology also contribute to define an animal's hearing abilities (Manley 1971; Echteler et al. 1994), we will only discuss the middle ear, due to space constraints.

## Measurements related to hearing sensitivity

This review will refer to studies that employed a variety of methods to measure the response of the auditory system to sound. These methods probe various points along the hearing pathway and can produce different results. It is, therefore, important to be aware of the main differences between the most frequently used measurement techniques in order to interpret their results appropriately.

## Behavior

Behavioral methods often involve training an animal to produce a motor output in response to a pure-tone acoustic stimulus (Heise 1953; Masterton et al. 1969; Konishi 1970). By varying the intensity and the frequency of the stimulus and recording the individual's response, threshold curves called audiograms are produced. Behavioral audiograms reveal the minimum intensity of sound at the ear that is necessary at each frequency to evoke the trained response. This method provides a natural assessment of hearing; it is noninvasive and it quantifies high-level neural

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responses to the stimuli. Although behavioral audiograms are commonly measured in mammals and birds (Masterton et al. 1969; Konishi 1970; Fay 1988; Dooling 2002), they are rarely obtained in amphibians and reptiles (Bulog and Schlegel 2000), which are notoriously difficult to train. The extent to which behavioral audiograms represent hearing abilities can be influenced by the effectiveness of the training regime.

### Electrophysiology

Neural activity may be recorded directly from various areas of the brain of animals, axons in the auditory (VIIIth) nerve, or from various locations in the inner ear, including hair cells. Electrodes can be prepared to obtain intra- or extracellular recordings from a single cell, or to record from an ensemble of cells (Kraus and McGee 1992; Eggermont and Ponton 2002). Hair cells in the main auditory sensory epithelium of all tetrapods are tonotopically organized, i.e., their frequency of peak sensitivity is a function of their position in the organ. This frequency-specific organization is conserved in the auditory nerve and in the midbrain. Assessments of the hearing range of animals through electrophysiological methods involving direct measurement from neurons, therefore, demand sampling the responses of enough cells to span the whole range of sensitivity. Such recordings tend to be in good agreement with behavioral audiograms (Konishi 1970; Dooling and Popper 2000). The neural response to sound can also be measured indirectly (without neuronal contact), through recordings of auditory-evoked responses in the brain, vestibulo-ocular reflexes and magnetic resonance imaging. Such methods are advantageous for being noninvasive, but they tend to produce results with higher thresholds than do measurements involving direct neuronal contact (Kraus and McGee 1992; Eggermont 2001; Eggermont and Ponton 2002; Todd et al. 2008). Another type of indirect technique is the recording of microphonic potentials using an electrode in the inner ear or on the round window. The combined electrical activity of the hair cells in response to mechanical excitation is then measured (Wever and Bray 1930, 1936; Tasaki et al. 1954).

### Eardrum vibration measurements

The frequency response of the eardrum vibration to acoustic stimuli has been assessed with various methods, including stroboscopic illumination (von Békésy 1960), capacitive probe (von Békésy 1960), the Mössbauer technique using a radioactive source (Hillman et al. 1964), displacement measurements

with laser interferometry (Khanna and Tonndorf 1972; Dragsten et al. 1974), and velocity measurements using laser Doppler vibrometry (Buunen and Vlaming 1981). Such measurements can be rapid and noninvasive, depending on the accessibility of the eardrum. While the spectral response of the eardrums to sound often coincides with the frequency ranges determined by behavioral tests or electrophysiology, it does not reflect signal losses that may occur in the ossicular chain (especially in cartilaginous middle ears), or during either signal transduction by the hair cells, or processing in the central nervous system.

### Variability and body size

The diversity of hearing sensitivity spectra has been summarized for various groups of vertebrates (Fay 1988, 1992; Dooling and Popper 2000; Dooling 2002). The extent of phylogenetic coverage found in the literature varies widely among classes, limiting comparisons of hearing performance among groups.

One major difference found between the middle ears of mammals and those of other tetrapods is that the ossicular chain of mammals is composed of three ossicles (malleus, incus, and stapes), whereas in nonmammals it comprises one ossicle (stapes or columella) and one cartilaginous element (extrastapes or extracolumella; Clack and Allin 2004). Until recently, the frequency ranges of hearing sensitivity in all nonmammals were believed to be restricted to below 12 kHz, whereas in mammals they often can extend beyond 40 kHz (Fay 1992). These differences, in addition to studies on single-ossicle ears showing transmission efficiency loss at high frequencies due to flexing of the extrastapes (Manley 1972a, 1972b, 1981), lead to a common belief that the single-ossicle design prevented nonmammals from evolving hearing sensitivity above 12 kHz (Manley and Gleich 1992). This hypothesis is currently being revisited as three species of Asian frogs (*Odorrana livida*, *Odorrana tormota* and *Huia cavitympanum*, Ranidae) have been shown to detect ultrasound, with efficient transmission of frequencies up to ~40 kHz by the middle ear and acoustically evoked potentials recorded in the midbrain up to 38 kHz (Feng et al. 2006; Arch et al. 2008, 2009; Gridi-Papp et al. 2008).

Body size has been reported to have a strong inverse correlation with the frequency of peak sensitivity both among and within species (Rosowski and Graybeal 1991; Dooling 1992; Hetherington 1992; Nummela 1995; Werner et al. 1998, 2002; Nummela et al. 1999; Dooling and Popper 2000;

Werner and Igc 2002; Gleich et al. 2005; Nummela and Sanchez-Villagra 2006). The sizes of the middle ear components and the volumes of the middle ear and mouth cavities scale with body size, resulting in correlated air compliances, resonances and, ultimately, middle ear tuning (Hetherington 1992; Rosowski 1994; Hemilä et al. 1995; Nummela 1995; Nummela et al. 1999; Huang et al. 2000; Nummela and Sanchez-Villagra 2006). All major tetrapod groups show, however, extensive interspecific variation around the average relation between body size and frequency of peak sensitivity. Evidence has been reported in several groups that such variation can be partly explained by adaptation to the physical or biotic environment (Okanoya and Dooling 1987, 1988; Dooling 1992; Wilczynski et al. 1993). Body size itself can be driven by environmental factors that affect development or select individuals for body size. Tetrapods, especially ectotherms, tend to be larger in colder environments, leading to latitudinal and altitudinal patterns of gradual change both across and within species (Narins and Smith 1986; Partridge and French 1996; Angilletta et al. 2004).

### The abiotic environment

Lewis and Fay (2004) provide an analytical discussion of the environmental factors that are most likely to have shaped hearing in tetrapods. We will focus on the empirical evidence of correlated variation between hearing sensitivity and environmental factors, principally medium characteristics, habitat acoustics, temperature, and noise.

### Medium characteristics

Tetrapod ears operate surrounded by a medium of air, water, or soil and the performance of the same ear in different media has been quantified in a few mammals and amphibians. Behavioral audiograms show that relative to air, human auditory thresholds increase underwater and these differences increase with frequency from 18 dB at 125 Hz to 56 dB at 8 kHz (Brandt and Hollien 1967). Marine mammals vary from hearing best in water (elephant seals, *Mirounga angustirostris*, Phocidae), in air (sea lions, *Zalophus californianus*, Otariidae), or hearing equally well in both media (harbor seals, *Phoca vitulina*, Phocidae; Richardson et al. 1995; Kastak and Schusterman 1998). In amphibians, airborne and underwater hearing have been compared in bullfrogs [*Lithobates catesbeianus*, formerly *Rana catesbeiana* (Ranidae)] using multi-unit acoustically evoked responses recorded in the torus semicircularis of the midbrain (Lombard et al. 1981). They revealed

underwater sensitivity to be equal or better than airborne sensitivity below 200 Hz, and up to 30 dB poorer above 200 Hz.

While comparing the hearing performance of a terrestrial species in air and underwater can reveal the effect of the change in medium, comparative analysis of the sensitivity and structure of specialists inhabiting each medium can point to the evolutionary changes involved in adaptation to that medium. All vertebrate inner ears are filled with fluid, which must be vibrated to stimulate the sensory hair cells (Lewis et al. 1985). The most obvious differences in structure and function exhibited by tetrapods specialized to each medium relate to the pathways taken by the acoustic input to the inner ear.

Air is a sound transmission medium with much lower impedance than the body tissues of vertebrates and therefore the direct transfer of acoustic signals from air into the tissues is minimal. Middle ears resolve the impedance mismatch between sound in air and the inner ear fluid, greatly increasing auditory sensitivity (Vlaming and Feenstra 1986; Jørgensen and Kannevorff 1998; Voss et al. 2000; Aibara et al. 2001; Koike et al. 2002; Stenfelt et al. 2002). The impedance matching is based on two features: (1) The large surface area ratio between the tympanic membrane (eardrum) which is in contact with the air, and the oval window, which contacts the inner ear fluid. (2) The vibration of the tympanic membrane is transferred to the oval window through one or two levers formed by the ossicular chain (two levers are found in amphibians, see Jørgensen and Kannevorff 1998; Mason and Narins 2002b). The problem of impedance mismatch, and therefore the importance of the middle ear, increases with body size and sound frequency. In small amphibians, external sound can cross the floor of the mouth and the tissues of the body wall to excite the air contained in the mouth and lungs (Wilczynski et al. 1987; Narins et al. 1988; Ehret et al. 1990, 1994; Hetherington and Lindquist 1999; Hetherington 2001; Mason 2006). The lungs and mouth connect to the middle ear cavity through the short and wide Eustachian tubes, that remain open at rest (but see Gridi-Papp et al. 2008), and provide an alternative acoustic pathway for auditory input (Narins et al. 1988). Middle ear structures have been secondarily lost in small species of several groups of amphibians, and this loss might be due to developmental reduction (Hetherington and Lindquist 1999). As an alternative explanation, small frogs tend to have relatively large lungs and thin body walls. Thus, their lungs might serve as an efficient conduit for sound pickup and delivery

to the middle ear, taking up the role of the eardrum in sound admission (Narins et al. 1988; Ehret et al. 1994). Such an hypothesis is supported by measurements showing that two species of frogs without eardrums have relatively low auditory thresholds (Hetherington 1992; Jaslow and Lombard 1996).

When sound propagates underwater, it passes into the body tissues of vertebrates with relatively little loss since the impedances of tissues and water are similar. The ear canal in whales and dolphins is narrow and plugged with dense cerumen (Clack 1992; Fritzsche 1992; Ketten 1997; Nummela et al. 2004) and it is believed to have lost its importance in conducting sound to the inner ears. High-frequency acoustic input in odontocetes is mainly through a channel of low-impedance fatty tissues that runs from the mid-line of the lower jaw to the temporal bones (Varanasi and Malins 1971; Norris 1980). The anatomical modifications found in the middle ears of secondarily aquatic nonmammalian vertebrates appear to be less extreme than those found in cetaceans, but a detailed mechanistic understanding of underwater hearing is still lacking in most cases (Hetherington 2008).

Fossorial animals live with their heads buried in sand or soil, which attenuates high-frequency sounds. Some fossorial insectivores, such as the African golden moles (Chrysochloridae) have evolved an extreme specialization for detection of low frequencies. The mallei in the middle ear are hypertrophied, and due to their increased mass and the fact that their center of mass is displaced from their rotatory axis, they vibrate out of phase with the skull in response to seismic vibrations, stimulating the inner ear (Mason and Narins 2001, 2002a; Willi et al. 2006b). In this mode, the relative motion between the malleus and the skull is due to the inertia of the malleus, rather than mechanical drive by the tympanic membrane.

Most terrestrial animals live with their ears surrounded by air, but with their feet on the ground. As the two media (air and ground) have different sound transmission properties, they can provide the animal with complementary input from the environment. For example, elephants are known to be highly sensitive to low-frequency sound (Heffner and Heffner 1980) and to produce and detect seismic signals (O'Connell et al. 1997; O'Connell-Rodwell et al. 2000, 2001). Seismic stimuli can be transmitted from the ground to the feet and up to the ears through bone conduction. The massive middle ear ossicles of elephants respond to vibration and stimulate the inner ears in a manner similar to that described above for golden moles

(Reuter et al. 1998). Among the tetrapods, seismic sensitivity is currently believed to be highest in amphibians (Ross and Smith 1978, 1979; Narins and Lewis 1984; Yu et al. 1991; Narins 2001). Besides tympanic hearing, amphibians possess a second middle ear mechanism: the opercularis system. It consists, in part, of a disk-shaped bone, the operculum, which abuts a portion of the oval window of the inner ear. The distal surface of the operculum receives the insertion of the opercularis muscle, which at its other end inserts into the cartilagenous suprascapula. This forms an alternative pathway for acoustic stimuli, in which seismic vibrations are transmitted from the ground to the shoulders through bone conduction, then through the tonic opercularis muscle to the operculum and into the inner ear (Hetherington 1985, 1987; Hetherington et al. 1986). Disruption of the opercularis system produces up to 6.5–13.1 dB loss in seismic sensitivity (Hetherington 1988).

### Acoustics of the habitat

The acoustics of the environment in which an animal lives and communicates can influence several aspects of sound transmission, potentially leading to adaptation of the animal's vocal and hearing systems (Richards and Wiley 1980; Penna and Solís 1996; Penna and Márquez 2007). Due to constraints of space, we will restrict this section to the well-studied case of rodents that live in tunnels, which is a clear example of repeated independent evolution of middle ear structure and function correlated with environmental acoustics.

Comparative studies in rodents have revealed a close relation between their habitat and features of their hearing, vocalizations, and anatomy (Heth et al. 1986; Heffner and Heffner 1992b, 1993; Heffner et al. 1994; Jackson et al. 1997; Lange et al. 2004, 2007; Begall and Burda 2006). Heth and colleagues investigated the acoustics in the tunnels used by blind mole rats (*Nannospalax ehrenbergi*, Spalacidae), and found that sound transmission was greatly decreased at high frequencies with best transmission at 440 Hz (Heth et al. 1986). Both the hearing range and the vocalizations of blind mole rats are shifted to low frequencies, presumably as an adaptation to the acoustics of the tunnels (Heth et al. 1986; Bruns et al. 1988). Later studies demonstrated that the patterns first observed in the blind mole rats may be generalized across rodents: subterranean species tend to have reduced sensitivity in comparison with surface dwellers, having higher sensitivity at low frequencies but much reduced

sensitivity at high frequencies. Species such as prairie dogs (genus *Cynomys*; Sciuridae), that are intermediate between subterranean and surface-dwelling rodents in the amount of time spent underground versus above the ground, also have intermediate sensitivity at low and at high frequencies. The hearing and vocal features of subterranean rodents are accompanied by modifications of the middle ear including: relatively larger eardrum lacking a pars flacida, enlarged incus, and reduced or missing middle ear muscles (Burda et al. 1992). In a recent study, Lange and colleagues (2007) evaluated the acoustics of tunnels used by subterranean rodents, and confirmed the patterns reported previously by Heth and colleagues (Heth et al. 1986), but they also found that some tunnels actually amplified sounds by up to 6.5 dB at frequencies between 200 and 800 Hz (Lange et al. 2007). They suggested that reduced peak sensitivity in subterranean rodents might be an adaptation to prevent overstimulation of the ears.

### Temperature

Gerhardt (1978) studied male advertisement calls and female acoustic preferences of two sympatric sister species of gray treefrogs (*Hyla versicolor* and *Hyla chrysoscelis*, Hylidae), in which the calls differ only in note rate. He found that increasing temperature caused males of both species to produce calls with higher note rates, and females of both species to prefer calls with higher note rates. The separation between the communication systems of the two species was therefore maintained because temperature biased both signalers and receivers in the same way. Walkowiak (1980) made multi-unit recordings in two species of frogs to assess the effect of temperature on midbrain auditory responses to pure tones. He found that sensitivity increased with temperature in both species, but stabilized at different optimal temperatures. Such optimal temperatures matched the temperatures in which the frogs most frequently call in nature, indicating that the hearing systems are adapted to the temperature range to which each of several frog species is naturally exposed. Nevertheless, studies of the responses of the eighth nerve to sound as a function of temperature in various frog species demonstrated that whereas the responses of the amphibian papillar fibers were highly temperature sensitive, responses of the basilar papillar fibers were strikingly temperature-insensitive (Stiebler and Narins 1990; van Dijk et al. 1990; Benedix et al. 1994).

### Noise

Abiotic noise produced by wind, flowing water, or rains typically has a broad spectrum with a low frequency peak. In response to abiotic background noise, hearing systems have been suggested to adapt by shifting their sensitivity ranges toward high frequencies (Langemann et al. 1998; Feng et al. 2006; Arch et al. 2009). One could also expect animals to evolve enhanced critical masking ratios at frequencies that are most biologically relevant to them, and in fact, parakeets, horseshoe bats, and green treefrogs have lowest critical masking ratios at the frequencies in which their absolute sensitivity is greatest, which correspond to the dominant frequencies of their calls (Dooling and Saunders 1975; Long 1977; Moss and Simmons 1986). This simple match is not a general pattern; however, as in most other mammals and birds that have been studied to date the critical masking ratio tends to simply increase with frequency (Dooling 1980; Okanoya and Dooling 1987; Klump 1996).

Besides their adverse effect in masking communication, natural abiotic acoustic sounds can also benefit animals by carrying relevant information about the environment. In the Namib Desert, substrate vibrations that are produced when the wind passes over vegetation-topped sand mounds result in seismic signals in the substrate that are detected at a distance by golden moles (Narins et al. 1997; Lewis et al. 2006). These animals cue in on spectral differences between the vibrations produced by the wind blowing over naked sand mounds versus over sand mounds topped with vegetation. By detecting wind-produced vibrations, the golden moles are able to localize and approach vegetation-covered mounds, within which their arthropod prey are most abundant. Many species of golden moles have extraordinarily hypertrophied mallei, which make them highly sensitive to seismic vibrations of the substrate (Mason and Narins 2002a; Mason 2003a, 2003b; Willi et al. 2006b).

### The biotic environment

Biotic factors include heterospecific, conspecific, or self-generated sounds to which the animal must be sensitive, and acoustic interference by sounds that other animals generate.

#### Heterospecific sounds

Cases of hearing sensitivity shaped by predator sounds have been reported in noctuid moths that rely on ultrasonic sensitivity to detect the echolocation calls of bats and initiate escape maneuvers

(Roeder 1962, 1966; Spangler 1988; Hoy 1992). The frequency ranges of sounds produced by predators and prey are also commonly found to match the hearing abilities of tetrapods. While many bats rely on echolocation to locate prey, some species cue in to the sounds produced by the insects or frogs that they consume. Echolocation calls are mostly ultrasonic and bats' ears are correspondingly tuned to high frequencies (Dalland 1965; Neuweiler 1984). Some bats present a second sensitivity peak at low frequencies (8–20 kHz). This secondary sensitivity peak has been suggested to be an adaptation to wide-range social communication, as audible frequencies transmit farther in the atmosphere than does ultrasound, or to locate prey items based on their sounds (Brown et al. 1978; Guppy and Coles 1988; Fuzessery et al. 1993; ter Hofstede et al. 2008). The frog-eating bat *Trachops cirrhosus* can be attracted to loudspeakers broadcasting low-frequency frog calls or pure tones. These bats' hearing sensitivity decreases from 15 to 5 kHz and increases again from 5 to 0.2 kHz, which is the range of most frog calls (Ryan et al. 1983).

Some of the highest acoustic sensitivities have been documented in nocturnal predators, which are greatly dependent on hearing to detect and seize prey (Neff and Hind 1955; Dalland 1965; Konishi 1973; van Dijk 1973; Neuweiler 1984; Dyson et al. 1998). High-frequency sensitivity in mammals has been suggested to be, in general, an adaptation for precise predator and prey localization based on vocalizations and wide-band sounds produced by animals moving on the substrate (Heffner and Heffner 1992a, 2008). The highly mobile external ears of most mammals are also believed to reflect the importance of detecting and locating heterospecifics (Flynn and Elliott 1965; Coles and Guppy 1986; Guppy and Coles 1988; Musicant et al. 1990).

### Conspecific sounds

When analyzed across species, the frequency of highest sensitivity of the auditory system often exhibits a close match with the dominant frequency of conspecific calls (Konishi 1970; Dooling et al. 1971; Dooling and Saunders 1975; Okanoya and Dooling 1988; Wilczynski et al. 1993). This pattern could be partly due to stabilizing selection by conspecifics, and partly by allometry, as both vocal fold length and middle ear size scale with body size and correlate inversely with the tuning of calls and ears.

When examined between the sexes, within a species, however, the frequency of best sensitivity does not always match the frequency of the calls.

In several species of frogs, the peak sensitivities obtained through recordings of evoked potentials in the midbrains of females were tuned to lower frequencies than the average male calls (Gerhardt and Doherty 1988; Ryan and Wilczynski 1988; Ryan 1990; Ryan and Rand 1990; Wilczynski et al. 2001). Such female midbrain recordings closely approximated the female's acoustic preferences. And several studies have found female acoustic preferences to be biased toward low frequency calls (Ryan 1980; Robertson 1986; Dyson and Passmore 1988; Gerhardt and Doherty 1988; Morris and Yoon 1989; Ryan et al. 1992). Female frogs, in these species, are larger than males, and this has been suggested to be the basis for sexual selection for large males: females, being larger than males, have their auditory systems tuned to lower frequencies than the average male call, and become maximally excited by the lowest pitched calls, which are produced by the largest males (Ryan et al. 1992).

Mismatches between auditory sensitivity and dominant song frequency have also been reported in house sparrows (e.g., Henry and Lucas 2008). The authors suggested that the mismatch in this species may reflect selection for high-frequency song and relaxed selection for a close match between sender and receiver due to small territory size.

### Self-produced sounds

Echolocating animals are unique in making intensive use of the vocal and auditory mechanisms in a self-matched manner (Konishi and Knudsen 1979; Au 1993; Grinnell 1995). Some species exhibit an extraordinary peak of auditory sensitivity at the echolocation call frequencies (Pollak et al. 1972). Such high sensitivity is not aimed at the echolocation calls themselves, but at the dim echos which reflect from the target objects. Synchronously with the emission of each echolocation call, the middle ear muscles contract to avoid overloading the ears before receiving the echos (Henson 1965; Suga et al. 1974).

Self-hearing is also important for vocal learning in birds and humans (Nottebohm 1972; Marler and Slabbekoorn 2004), and for stimulating ovulation in ring doves (Cheng 1992). The frequency content of these sounds overlaps, however, with that of sounds produced by conspecifics during communication. It seems, therefore, unlikely that the need to hear self would generate any unique selective pressure on hearing frequency sensitivity that could lead to specialization.

### Biotic noise

Biotic noise is often produced by a large number of individuals emitting advertisement calls or making sound as they move. It tends to have a narrower spectrum than abiotic noise sources such as wind or rain.

In multi-species breeding assemblages, acoustic communication may be hindered by the noise produced by heterospecific advertisement calls. Several studies have described interspecific acoustic variation in such assemblages as acoustic resource partitioning, but few of them have actually attempted to test the hypothesis underlying this ecological term, i.e., verifying if the distribution of acoustic traits is more regular than expected by chance (Duellman and Pyles 1983; Heller and Helversen 1989; Kingston et al. 2000; Chek et al. 2003). In addition, the existing evidence is based on advertisement calls and does not address hearing. It is tempting to consider, however, that partitioning of the acoustic space for calls should also lead to adaptation of hearing, as the frequency ranges of advertisement calls and hearing tend to be highly correlated when compared across species (Konishi 1970; Dooling et al. 1971; Dooling and Saunders 1975; Okanoya and Dooling 1988; Wilczynski et al. 1993).

Additional evidence is available from studies of reproductive character displacement in advertisement calls. In frogs, these calls are frequently the main or the only species-isolation mechanism between two species, and in some cases the acoustic structure of the advertisement calls is more divergent in the zone of contact than in allopatry (Littlejohn 1965; Blair 1974; Fouquette 1975). Several studies have also assessed the acoustic preferences of the females, and found them to favor the divergence of the calls in sympatry (Gerhardt 1994; Márquez and Bosch 1997; Pfennig 2000; Höbel and Gerhardt 2003; Pfennig and Ryan 2007). When the fitness of the hybrids is reduced, female preferences should be selected to avoid hybrid matings. Hearing sensitivity spectrum could be involved in changes in female preferences for the frequencies of male calls (Capranica and Moffat 1983) although the available examples of acoustic character displacement (cited above) only involve divergence of temporal characters of the call.

### Multiple sensitivity peaks

While most tetrapods exhibit a single peak with hearing sensitivity gradually rolling off at higher and lower frequencies, some animals exhibit multiple sensitivity peaks, which can reflect different

ecological pressures. Several species of bats have been found to exhibit two hearing sensitivity peaks the higher one matching the frequency range of the echolocation calls and the lower one matching the frequency range of social signals or sounds produced by prey (see Biotic environment: Heterospecific sounds).

Ears that are selected to respond with high sensitivity to more than one frequency range can be especially informative about limitations and versatility of the auditory system. As mentioned before (Abiotic factors: Noise), golden moles have hypertrophied mallei and are highly sensitive to low-frequency seismic vibrations. These animals produce vocalizations with fundamental frequencies  $\sim 1\text{--}2.2$  kHz and considerable energy up to 20 kHz in some calls (Willi et al. 2006a). While enlargement of the malleus is a key aspect of their seismic sensitivity, such an adaptation could have compromised acoustic hearing, since the massive mallei should resist vibration at high frequencies. As an ingenious solution to this problem, the mallei of golden moles rotate around different axes in response to acoustic or seismic stimuli (Willi et al. 2006a). The large mass of the malleus is centered in relation to the acoustic rotation axis but it remains displaced relative to the seismic rotation axis. This arrangement allows for high seismic sensitivity with minimal loss of acoustic sensitivity.

Studies of auditory sensitivity in frogs, based on midbrain auditory-evoked potentials, often reveal two sensitivity peaks (Mudry and Capranica 1987; Wilczynski et al. 1993, 2001). The double sensitivity in this case, however, is believed not to reflect multiple ecological selective pressures, but multiple hearing organs. The low-frequency peak corresponds to the sensitivity range of the amphibian papilla whereas the high-frequency peak corresponds to the sensitivity of the basilar papilla (Capranica and Moffat 1983). It has been suggested that low frequencies excite the ear predominantly via the extratympanic route and that high-frequencies preferentially enter the ear through the tympanic route thereby exciting the basilar papilla (Wilczynski et al. 1987).

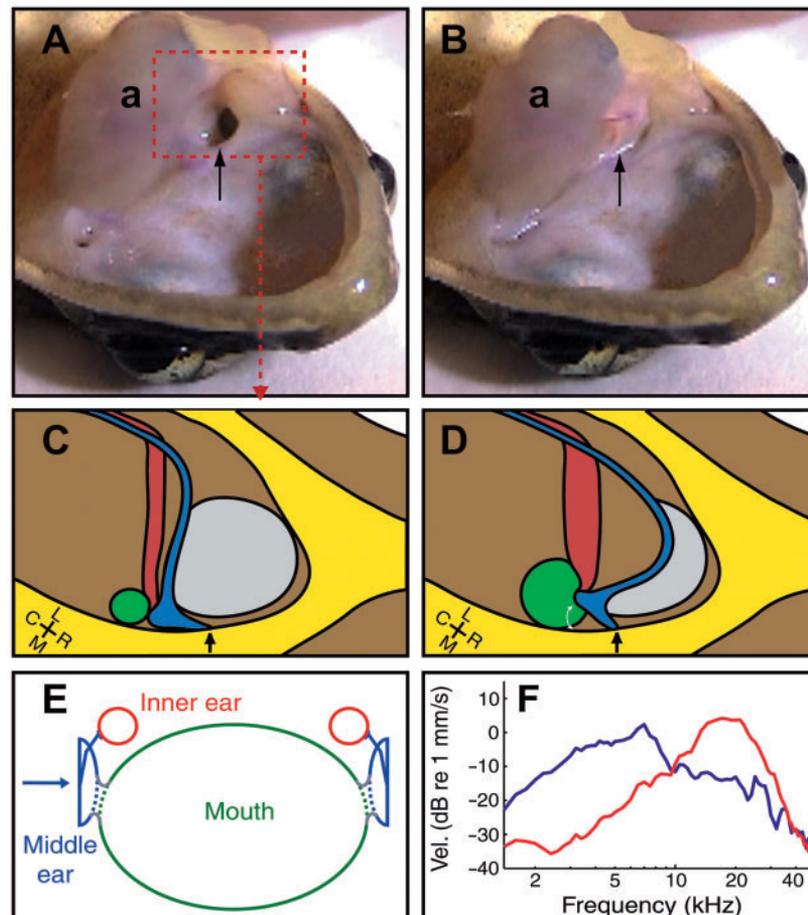
### Dynamic middle ear tuning

Besides evolving changes to the structure of the middle ear, vertebrates can behaviorally adjust their peripheral auditory mechanics according to environmental constraints. The best known behavioral mechanism of this type is the acoustic reflex, in which the middle ear muscles contract to restrain the movement of the middle ear elements

(Borg 1972; Nuttall 1974; Oeckinghaus and Schwartzkopff 1983). The acoustic reflex produces extensive attenuation at low frequencies and little or no gain at high frequencies. It is triggered by intense external sound, but similar contractions of the middle ear muscles can be produced in the absence of external sound, by command from the central nervous system, in synchrony with vocalizations (Suga et al. 1974; Borg and Zakrisson 1975).

In addition to direct muscular action on the middle ear elements, a novel mechanism has been recently described (Gridi-Papp et al. 2008).

Compared with mammals, most frogs possess large tympanic membranes and compressed middle ear cavities which connect to the mouth through short and wide Eustachian tubes. Such a connection was believed to remain permanently open, but the Chinese concave-eared torrent frog (*O. tormota*) has been shown to actively close its Eustachian tubes (Gridi-Papp et al. 2008). The isolation of the middle ear cavity significantly diminishes its compliance, producing extensive attenuation ( $\sim 26$  dB) of low frequencies (Fig. 1). With the tiny middle ear cavity isolated from the mouth, the resonant



**Fig. 1** Active Eustachian tube (ET) closure shifts middle ear tuning in the frog *O. tormota*. (A) Awake male with the mouth open, showing the wide ET opening (arrow) in the roof of the mouth; “a” labels the tongue. (B) The same male seconds after A, with the ET fully closed. (C) Schematic representation of the anatomy within the hashed red rectangle in A, depicting the anatomy involved in ET closure; the perimeter of the ET (gray) is delimited by the skull (yellow) rostrally, laterally and medially, but the caudal perimeter is delimited by the flexible anterior hyoid horn (blue), which has a narrow attachment to the skull (arrow); the submaxillary muscle (red) inserts in the caudal thickening of the hyoid near its attachment to the skull, next to the petrohyoid muscles (green), which form an angle with the plane of the hyoid cartilage. Coordinates: R=rostral, C=caudal, M=medial, L=lateral. (D) Contraction of the submaxillary muscle causes the hyoid cartilage to bend and pivot on its attachment to the skull, closing the ET. The movement is enhanced with pressure exerted by the petrohyoid muscles during contraction. (E) ET closure isolates the middle ear cavity from the mouth cavity, drastically reducing the volume of air and the compliance of the cavity behind the eardrums (blue arrow). (F) Vibration velocity of the TM in response to acoustic stimulation by a pure tone at various frequencies. In relation to the resting condition with the ET open (blue curve), closure of the ET (red curve) produces up to 26 dB attenuation below 10 kHz and up to 20 dB gain above 10 kHz. (Figure modified from Gridi-Papp et al. 2008, with permission. Copyright 2008 National Academy of Sciences, USA).

properties of the ear are altered and a gain of up to 20 dB is produced at high frequencies.

### Diversity and evolution

Phylogenetic history explains significant portions of the diversity observed in the structure and the hearing sensitivity spectra of the middle ear of tetrapods. Middle and external ears evolved late after the initial vertebrate colonization of land and it occurred multiple times, thereby conferring structural differences among tetrapods (Clack and Allin 2004). Vertebrate classes also exhibit important differences in the mechanics and organization of the sensory epithelia of the inner ear (Manley and Clack 2004). These differences are maintained across most modern taxa and they can potentially bias the evolutionary response of species to environmental selection pressures (i.e., represent evolutionary constraint *sensu* Maynard-Smith et al. 1985).

This review found evidence of various environmental transitions that relate to changes in hearing ranges or in middle ear structure in tetrapods. Both phylogeny and environment should, therefore, be considered in a comprehensive explanation of the diversity in tetrapod hearing. The contributions of phylogeny and environment to hearing diversity can be estimated quantitatively, through the analysis of pools of taxa that represent independent transitions between environmental conditions (Felsenstein 1985).

### Conclusions

The number of species for which data on hearing performance and middle ear mechanics is available is still small. The extent of taxonomic sampling is variable among vertebrate groups and widely diverging designs have been found. The generality of some patterns of environmental effects on hearing can already be established, but several others should emerge with the continued publication of measurements on additional species.

Body size is inversely correlated with frequency range of hearing and directly with the size of the middle ear structures in all major tetrapod groups. Such relationships should not be seen as confounding factors to be removed from the analysis of environmental effects, because body size itself is frequently driven by abiotic or biotic environmental factors. Such environmental factors can, therefore, alter the hearing of animals via changes in body size.

Tetrapods inhabit media (air, water, or underground) that differ drastically in impedance and sound transmission properties, factors that affect

hearing sensitivity and the frequency range of detectable stimuli. Various vertebrate groups exhibit anatomical specializations of the middle ear that relate to coupling of the stimulus to the inner ear or to frequency range. Such specializations appear to represent adaptations for enhanced stimulus detection.

The current literature also provides various levels of support for the influence of biotic environmental factors on hearing frequency range. Such factors include interactions among species, within species and even the perception of self-produced signals reflected by the environment. Hearing systems appear to adapt both when individuals are selected to better detect a given signal, or when they are selected to cope with acoustic interference.

Two mechanisms of behavioral middle ear tuning have been described, both with suggested protective roles, one involving muscular restraint of the ossicular chain, and the other involving an increase of the impedance of the air cavity behind the eardrum.

As measurements accumulate on hearing spectrum, middle ear morphology and acoustic ecology of tetrapods, it becomes feasible to estimate the proportion of variation in middle ear structure and hearing range that can be associated with a given environmental factor.

Accelerating the development of knowledge on the relations between vertebrate hearing and the environment is a pressing necessity, as fast technological developments are resulting in rapid alteration of the acoustics in most environments and potentially risking the health of humans and animals. The study of hearing structure and function in additional taxa will expand the comparative basis of evidence and add power to the analysis of environmental effects on hearing. Such taxonomic sampling should continue to reveal unexpected novel mechanisms and extreme adaptations to environmental factors. In depth studies of such extreme adaptations may reveal general principles not readily observed in less extreme species, or in the words of the Danish Nobel laureate August Krogh: "For many problems, there will be an animal for which it can be most conveniently studied". The study of additional species should, therefore, be intensified until most vertebrate groups, geographic areas and environmental factors are sampled.

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## References

- Aibara R, Welsh JT, Puria S, Goode RL. 2001. Human middle-ear sound transfer function and cochlear input impedance. *Hear Res* 152:100–9.
- Angilletta MJ, Steury TD, Sears MW. 2004. Temperature, growth rate, and body size in ectotherms: fitting pieces of a life-history puzzle. *Integr Comp Biol* 44:498–509.
- Arch VS, Grafe TU, Gridi-Papp M, Narins PM. 2009. Pure ultrasonic communication in an endemic Bornean frog. *PLoS ONE* 4:e5413.
- Arch VS, Grafe TU, Narins PM. 2008. Ultrasonic signaling by a Bornean frog. *Biol Lett* 4:19–22.
- Au WWL. 1993. *The Sonar of Dolphins*. New York: Springer.
- Begall S, Burda H. 2006. Acoustic communication and burrow acoustics are reflected in the ear morphology of the coruro (*Spalacopus cyanus*, Octodontidae), a social fossorial rodent. *J Morphol* 267:382–90.
- Benedix JH Jr, Pedemonte M, Velluti R, Narins PM. 1994. Temperature-dependence of two-tone rate suppression in the northern leopard frog *Rana pipiens pipiens*. *J Acoust Soc Am* 96:2738–45.
- Blair WF. 1974. Character displacement in frogs. *Am Zool* 14:1119–25.
- Borg E. 1972. On the change in the acoustic impedance of the ear as a measure of middle ear muscle reflex activity. *Acta Otolaryngol* 74:163–71.
- Borg E, Zakrisson JE. 1975. The activity of the stapedius muscle in man during vocalization. *Acta Otolaryngol* 79:325–33.
- Brandt JF, Hollien H. 1967. Underwater hearing thresholds in man. *J Acoust Soc Am* 42:966–71.
- Brown PE, Grinnell AD, Harrison JB. 1978. The development of hearing in the pallid bat, *Antrozous pallidus*. *J Comp Physiol* 126:169–82.
- Bruns V, Müller M, Hofer W, Heth G, Nevo E. 1988. Inner ear structure and electrophysiological audiograms of the subterranean mole rat, *Spalax ehrenbergi*. *Hear Res* 33:1–9.
- Bulog B, Schlegel P. 2000. Functional morphology of the inner ear and underwater audiograms of *Proteus anguinus* (Amphibia, Urodela). *Pflügers Arch* 439:r165–7.
- Burda H, Bruns V, Hickman GC. 1992. The ear in subterranean Insectivora and Rodentia in comparison with ground-dwelling representatives. I. Sound conducting system of the middle ear. *J Morphol* 214:49–61.
- Buunen TJJ, Vlaming MSMG. 1981. Laser-Doppler velocity meter applied to tympanic membrane vibrations in cat. *J Acoust Soc Am* 69:744–50.
- Capranica RR, Moffat AJM. 1983. Neurobehavioral correlates of sound communication in anurans. In: Ewert J, Capranica RR, Ingle DJ, editors. *Advances in vertebrate neuroethology*. New York: Plenum Press. p. 710–30.
- Chek AA, Bogart JP, Lougheed SC. 2003. Mating signal partitioning in multi-species assemblages: a null model test using frogs. *Ecol Lett* 6:235–47.
- Cheng M. 1992. For whom does the female dove coo? A case for the role of vocal self-stimulation. *Anim Behav* 43:1035–44.
- Clack JA. 1992. The stapes of *Acanthostega gunnari* and the role of the stapes in early tetrapods. In: Webster DB, Fay RR, Popper AN, editors. *The evolutionary biology of hearing*. New York: Springer. p. 405–20.
- Clack JA, Allin E. 2004. The evolution of single- and multiple-ossicle ears in fishes and tetrapods. In: Manley GA, Popper AN, Fay RR, editors. *Evolution of the vertebrate auditory system*. New York: Springer. p. 128–63.
- Coles RB, Guppy A. 1986. Biophysical aspects of directional hearing in the Tammar wallaby, *Macropus eugenii*. *J Exp Biol* 121:371–94.
- Dalland JI. 1965. Hearing sensitivity in bats. *Science* 150:1185–6.
- Dooling RJ. 1980. Behavior and psychophysics of hearing in birds. In: Popper AN, Fay RR, editors. *Comparative studies of hearing in vertebrates*. New York: Springer. p. 261–88.
- Dooling RJ. 1992. Hearing in birds. In: Webster DB, Fay RR, Popper AN, editors. *The evolutionary biology of hearing*. New York: Springer. p. 545–60.
- Dooling RJ. 2002. Avian hearing and the avoidance of wind turbines. Golden, Colorado: National Renewable Energy Laboratory.
- Dooling RJ, Fay RR, Popper AN. 2000. *Comparative hearing: birds and reptiles*. New York: Springer.
- Dooling RJ, Mulligan JA, Miller JD. 1971. Auditory sensitivity and song spectrum of the common canary (*Serinus canarius*). *J Acoust Soc Am* 50:700–9.
- Dooling RJ, Popper AN. 2000. Hearing in birds and reptiles: an overview. In: Dooling RJ, Fay RR, Popper AN, editors. *Comparative hearing: birds and reptiles*. New York: Springer. p. 308–59.
- Dooling RJ, Saunders JC. 1975. Hearing in the parakeet (*Melopsittacus undulatus*): absolute thresholds, critical ratios, frequency difference limens, and vocalizations. *J Comp Physiol Psychol* 88:1–20.
- Dragsten PR, Webb WW, Paton JA, Capranica RR. 1974. Auditory membrane vibrations: measurements at sub-angstrom levels by optical heterodyne spectroscopy. *Science* 185:55–7.
- Duellman WE, Pyles RA. 1983. Acoustic resource partitioning in anuran communities. *Copeia* 1983:639–49.
- Dusenbery DB. 1992. *Sensory ecology*. New York: W.H. Freeman.
- Dyson ML, Klump GM, Gauger B. 1998. Absolute hearing thresholds and critical masking ratios in the European

- barn owl: a comparison with other owls. *J Comp Physiol* 182:695–702.
- Dyson ML, Passmore NI. 1988. Two-choice phonotaxis in *Hyperolius marmoratus* (Anura: Hyperoliidae): the effect of temporal variation in presented stimuli. *Anim Behav* 36:648–52.
- Echteler SM, Fay RR, Popper AN. 1994. Structure of the mammalian cochlea. In: *Comparative hearing: mammals*. New York: Springer. p. 134–71.
- Eggermont JJ. 2001. Between sound and perception: reviewing the search for a neural code. *Hear Res* 157:1–42.
- Eggermont JJ, Ponton CW. 2002. The neurophysiology of auditory perception: from single units to evoked potentials. *Audiol Neurootol* 7:71–99.
- Ehret G, Keilwerth E, Kamada T. 1994. The lung-eardrum pathway in three treefrog and four dendrobatid frog species: some properties of sound transmission. *J Exp Biol* 195:329–43.
- Ehret G, Tautz J, Schmitz B, Narins PM. 1990. Hearing through the lungs: lung-eardrum transmission of sound in the frog *Eleutherodactylus coqui*. *Naturwissenschaften* 77:192–4.
- Fay RR. 1988. *Hearing in vertebrates: a psychoacoustics databook*. Winnetka, IL: Hill-Fay Associates.
- Fay RR. 1992. Structure and function in sound discrimination among vertebrates. In: Webster DB, Fay RR, Popper AN, editors. *The evolutionary biology of hearing*. New York: Springer. p. 229–63.
- Fay RR, Popper AN. 1994. *Comparative hearing: mammals*. New York: Springer.
- Fay RR, Popper AN. 1998. *Comparative hearing: fish and amphibians*. New York: Springer.
- Felsenstein J. 1985. Phylogenies and the comparative method. *Am Nat* 125:1–15.
- Feng AS, Narins PM, Xu C, Lin W, Yu Z, Qiu Q, Xu Z, Shen J. 2006. Ultrasonic communication in frogs. *Nature* 440:333–6.
- Flynn WE, Elliott DN. 1965. Role of the pinna in hearing. *J Acoust Soc Am* 38:104–105.
- Fouquette MJ. 1975. Speciation in chorus frogs. I. Reproductive character displacement in the *Pseudacris nigrita* complex. *Syst Zool* 24:16–23.
- Fritsch B. 1992. The water-to-land transition evolution of the tetrapod basilar papilla middle ear and auditory nuclei. In: Webster DB, Fay RR, Popper AN, editors. *The evolutionary biology of hearing*. New York: Springer. p. 351–76.
- Fuzessery ZM, Buttenhoff P, Andrews B, Kennedy JM. 1993. Passive sound localization of prey by the pallid bat (*Antrozous p. pallidus*). *J Comp Physiol* 171:767–77.
- Gerhardt HC. 1978. Temperature coupling in the vocal communication system of the gray tree frog, *Hyla versicolor*. *Science* 199:992–4.
- Gerhardt HC. 1994. Reproductive character displacement of female mate choice in the grey treefrog, *Hyla chrysoscelis*. *Anim Behav* 47:959–69.
- Gerhardt HC, Doherty JA. 1988. Acoustic communication in the gray treefrog, *Hyla versicolor*: evolutionary and neurobiological implications. *J Comp Physiol* 162:261–78.
- Gleich O, Dooling RJ, Manley GA. 2005. Audiogram, body mass, and basilar papilla length: correlations in birds and predictions for extinct archosaurs. *Naturwissenschaften* 92:595–8.
- Gridi-Papp M, Feng AS, Shen J, Yu Z, Rosowski JJ, Narins PM. 2008. Active control of ultrasonic hearing in frogs. *Proc Natl Acad Sci USA* 105:11014–9.
- Grinnell AD. 1995. Hearing in bats: an overview. In: Popper AN, Fay RR, editors. *Hearing by bats*. New York: Springer. p. 1–36.
- Guppy A, Coles RB. 1988. Acoustical and neural aspects of hearing in the Australian gleaning bats, *Macroderma gigas* and *Nyctophilus gouldi*. *J Comp Physiol* 162:653–68.
- Heffner HE, Heffner RS. 2008. High-frequency hearing. In: Basbaum AI, Kaneko A, Shepherd GM, Westheimer G, editors. *The senses: a comprehensive reference*. New York: Academic Press. p. 55–60.
- Heffner RS, Heffner HE. 1980. Hearing in the elephant (*Elephas maximus*). *Science* 208:518–20.
- Heffner RS, Heffner HE. 1992a. Evolution of sound localization in mammals. In: *The evolutionary biology of hearing*. New York: Springer. p. 691–715.
- Heffner RS, Heffner HE. 1992b. Hearing and sound localization in blind mole rats (*Spalax ehrenbergi*). *Hear Res* 62:206–16.
- Heffner RS, Heffner HE. 1993. Degenerate hearing and sound localization in naked mole rats (*Heterocephalus glaber*), with an overview of central auditory structures. *J Comp Neurol* 331:418–33.
- Heffner RS, Heffner HE, Contos C, Kearns D. 1994. Hearing in prairie dogs: transition between surface and subterranean rodents. *Hear Res* 73:185–9.
- Heise GA. 1953. Auditory thresholds in the pigeon. *Am J Psychol* 66:1–19.
- Heller K, Helversen OV. 1989. Resource partitioning of sonar frequency bands in rhinolophoid bats. *Oecologia* 80:178–86.
- Hemilä S, Nummela S, Reuter T. 1995. What middle ear parameters tell about impedance matching and high frequency hearing. *Hear Res* 85:31–44.
- Henson OWJ. 1965. The activity and function of the middle-ear muscles in echo-locating bats. *J Physiol* 180:871–87.
- Henry KS, Lucas JR. 2008. Coevolution of auditory sensitivity and temporal resolution with acoustic signal space in three songbirds. *Anim Behav* 76:1659–71.
- Hetherington TE. 1985. Role of the opercularis muscle in seismic sensitivity in the bullfrog, *Rana catesbeiana*. *J Exp Zool* 235:27–34.
- Hetherington TE. 1988. Biomechanics of vibration reception in the bullfrog, *Rana catesbeiana*. *J Comp Physiol* 163:43–52.
- Hetherington TE. 1987. Physiological features of the opercularis muscle and their effects on vibration sensitivity in the bullfrog *Rana catesbeiana*. *J Exp Biol* 131:189–204.

- Hetherington TE. 1992. The effects of body size on the evolution of the amphibian middle ear. In: Webster DB, Fay RR, Popper AN, editors. The evolutionary biology of hearing. New York: Springer. p. 421–38.
- Hetherington TE. 2001. Laser vibrometric studies of sound-induced motion of the body walls and lungs of salamanders and lizards: implications for lung-based hearing. *J Comp Physiol* 187:499–507.
- Hetherington TE. 2008. Comparative anatomy and function of hearing in aquatic amphibians, reptiles, and birds. In: Thewissen JGM, Nummela S, editors. Sensory evolution on the threshold: adaptations in secondarily aquatic vertebrates. Berkeley: University of California Press. p. 183–209.
- Hetherington TE, Jaslow AP, Lombard RE. 1986. Comparative morphology of the amphibian opercularis system: I. General design features and functional interpretation. *J Morphol* 190:43–61.
- Hetherington TE, Lindquist ED. 1999. Lung-based hearing in an “earless” anuran amphibian. *J Comp Physiol* 184:395–401.
- Heth G, Frankenberg E, Nevo E. 1986. Adaptive optimal sound for vocal communication in tunnels of a subterranean mammal (*Spalax ehrenbergi*). *Cell Mol Life Sci* 42:1287–9.
- Hillman P, Schechter H, Rubinstein M. 1964. Application of the Mossbauer technique to the measurement of small vibrations in the ear. *Rev Mod Phys* 36:360.
- Höbel G, Gerhardt HC. 2003. Reproductive character displacement in the acoustic communication system of green tree frogs (*Hyla cinerea*). *Evolution* 57:894–904.
- Hoy RR. 1992. The evolution of hearing in insects as an adaptation to predation from bats. In: Webster DB, Fay RR, Popper AN, editors. The evolutionary biology of hearing. New York: Springer. p. 115–29.
- Huang GT, Rosowski JJ, Peake WT. 2000. Relating middle-ear acoustic performance to body size in the cat family: measurements and models. *J Comp Physiol* 186:447–65.
- Jackson LL, Heffner HE, Heffner RS. 1997. Audiogram of the fox squirrel (*Sciurus niger*). *J Comp Psychol* 111:100–4.
- Jaslow AP, Lombard RE. 1996. Hearing in the neotropical frog, *Atelopus chiriquiensis*. *Copeia* 1996:428–32.
- Jørgensen MB, Kannevorf M. 1998. Middle ear transmission in the grass frog, *Rana temporaria*. *J Comp Physiol* 182:59–64.
- Kastak D, Schusterman RJ. 1998. Low-frequency amphibious hearing in pinnipeds: methods, measurements, noise, and ecology. *J Acoust Soc Am* 103:2216–28.
- Ketten DR. 1997. Structure and function in whale ears. *Bioacoustics* 8:103–35.
- Khanna SM, Tonndorf J. 1972. Tympanic membrane vibrations in cats studied by time-averaged holography. *J Acoust Soc Am* 51:1904–20.
- Kingston T, Jones G, Zubaid A, Kunz TH. 2000. Resource partitioning in rhinolophoid bats revisited. *Oecologia* 124:332–42.
- Klump GM. 1996. Bird communication in the noisy world. In: Kroodsma DE, Miller EH, editors. Ecology and evolution of acoustic communication in birds. Ithaca, NY: Cornell University Press. p. 321–38.
- Koike T, Wada H, Kobayashi T. 2002. Modeling of the human middle ear using the finite-element method. *J Acoust Soc Am* 111:1306–17.
- Konishi M. 1970. Comparative neurophysiological studies of hearing and vocalizations in songbirds. *J Comp Physiol* 66:257–72.
- Konishi M. 1973. How the owl tracks its prey. *Am Sci* 61:414–74.
- Konishi M, Knudsen EI. 1979. The oilbird: hearing and echolocation. *Science* 204:425–7.
- Kraus N, McGee T. 1992. Electrophysiology of the human auditory system. In: Popper AN, Fay RR, editors. The mammalian auditory pathway: neurophysiology. New York: Springer. p. 335–403.
- Lange S, Burda H, Wegner R, Dammann P, Begall S, Kawalika M. 2007. Living in a “stethoscope”: burrow-acoustics promote auditory specializations in subterranean rodents. *Naturwissenschaften* 94:134–8.
- Lange S, Stalleicken J, Burda H. 2004. Functional morphology of the ear in fossorial rodents, *Microtus arvalis* and *Arvicola terrestris*. *J Morphol* 262:770–9.
- Langemann U, Gauger B, Klump GM. 1998. Auditory sensitivity in the great tit: perception of signals in the presence and absence of noise. *Anim Behav* 56:763–9.
- Lewis ER, Fay RR. 2004. Environmental variables and the fundamental nature of hearing. In: Manley GA, Popper AN, Fay RR, editors. Evolution of the vertebrate auditory system. New York: Springer. p. 27–54.
- Lewis ER, Leverenz EL, Bialek WS. 1985. The vertebrate inner ear. Boca Raton, FL: CRC Press.
- Lewis ER, Narins PM, Jarvis JUM, Bronner G, Mason MJ. 2006. Preliminary evidence for the use of microseismic cues for navigation by the Namib golden mole. *J Acoust Soc Am* 119:1260–8.
- Littlejohn MJ. 1965. Premating isolation in the *Hyla ewingi* complex (Anura: Hylidae). *Evolution* 19:234–43.
- Lombard RE, Fay RR, Werner YL. 1981. Underwater hearing in the frog, *Rana catesbeiana*. *J Exp Biol* 91:57–71.
- Long GR. 1977. Masked auditory thresholds from the bat, *Rhinolophus ferrumequinum*. *J Comp Physiol* 116:247–55.
- Manley GA. 1971. Some aspects of the evolution of hearing in vertebrates. *Nature* 230:506–9.
- Manley GA. 1972a. Frequency response of the ear of the Tokay gecko. *J Exp Zool* 181:159–68.
- Manley GA. 1972b. The middle ear of the Tokay Gecko. *J Comp Physiol* 81:239–50.
- Manley GA. 1981. A review of the auditory physiology of the reptiles. *Progr Sens Physiol* 2:49–134.
- Manley GA, Clack JA. 2004. An outline of the evolution of vertebrate hearing organs. In: Manley GA, Popper AN, Fay RR, editors. Evolution of the vertebrate auditory system. New York: Springer. p. 1–26.

- Manley GA, Gleich O. 1992. Evolution and specialization of function in the avian auditory periphery. In: Webster DB, Fay RR, Popper AN, editors. *The evolutionary biology of hearing*. New York: Springer. p. 561–80.
- Marler P, Slabbekoorn HW. 2004. *Nature's music: the science of birdsong*. San Diego: Academic Press.
- Márquez R, Bosch J. 1997. Male advertisement call and female preference in sympatric and allopatric midwife toads. *Anim Behav* 54:1333–45.
- Mason MJ. 2003a. Bone conduction and seismic sensitivity in golden moles (Chrysochloridae). *J Zool* 260:405–13.
- Mason MJ. 2003b. Morphology of the middle ear of golden moles (Chrysochloridae). *J Zool* 260:391–403.
- Mason MJ. 2006. Pathways for sound transmission to the inner ear in amphibians. In: Narins PM, Feng AS, Fay RR, editors. *Hearing and sound communication in amphibians*. New York: Springer. p. 147–83.
- Mason MJ, Narins PM. 2001. Seismic signal use by fossorial mammals. *Am Zool* 41:1171–84.
- Mason MJ, Narins PM. 2002a. Seismic sensitivity in the desert golden mole (*Eremitalpa granti*): a review. *J Comp Psychol* 116:158–63.
- Mason MJ, Narins PM. 2002b. Vibrometric studies of the middle ear of the bullfrog *Rana catesbeiana* I. The extrastapes. *J Exp Biol* 205:3153–65.
- Masterton B, Heffner H, Ravizza R. 1969. The evolution of human hearing. *J Acoust Soc Am* 45:966–85.
- Maynard-Smith J, Burian R, Kauffman S, Alberch P, Campbell J, Goodwin B, Lande R, Raup D, Wolpert L. 1985. Developmental constraints and evolution. *Q Rev Biol* 60:265–87.
- Morris MR, Yoon SL. 1989. A mechanism for female choice of large males in the treefrog *Hyla chrysoscelis*. *Behav Ecol Sociobiol* 25:65–71.
- Moss CF, Simmons AM. 1986. Frequency selectivity of hearing in the green treefrog, *Hyla cinerea*. *J Comp Physiol* 159:257–66.
- Mudry KM, Capranica RR. 1987. Correlation between auditory thalamic area evoked responses and species-specific call characteristics. *J Comp Physiol* 161:407–16.
- Musicant AD, Chan JCK, Hind JE. 1990. Direction-dependent spectral properties of cat external ear: new data and cross-species comparisons. *J Acoust Soc Am* 87:757–81.
- Narins PM. 2001. Vibration communication in vertebrates. In: Barth F, Schmidt A, editors. *Ecology of sensing*. New York: Springer. p. 127–48.
- Narins PM, Ehret G, Tautz J. 1988. Accessory pathway for sound transfer in a neotropical frog. *Proc Natl Acad Sci USA* 85:1508–12.
- Narins PM, Lewis ER. 1984. The vertebrate ear as an exquisite seismic sensor. *J Acoust Soc Am* 76:1384–7.
- Narins PM, Lewis ER, Jarvis JJUM, O'Riain J. 1997. The use of seismic signals by fossorial Southern African mammals: a neuroethological gold mine. *Brain Res Bull* 44:641–6.
- Narins PM, Smith SL. 1986. Clinal variation in anuran advertisement calls: basis for acoustic isolation? *Behav Ecol Sociobiol* 19:135–41.
- Neff WD, Hind JE. 1955. Auditory thresholds of the cat. *J Acoust Soc Am* 27:480–3.
- Neuweiler G. 1984. Foraging, echolocation and audition in bats. *Naturwissenschaften* 71:446–55.
- Norris KS. 1980. Peripheral sound processing in odontocetes. In: Busnel RG, Fish JF, editors. *Animal sonar systems*. New York: Plenum Publishing Co. p. 495–509.
- Nottebohm F. 1972. The origins of vocal learning. *Am Nat* 106:116–40.
- Nummela S. 1995. Scaling of the mammalian middle ear. *Hear Res* 85:18–30.
- Nummela S, Sanchez-Villagra MR. 2006. Scaling of the marsupial middle ear and its functional significance. *J Zool* 270:256–67.
- Nummela S, Thewissen JGM, Bajpai S, Hussain ST, Kumar K. 2004. Eocene evolution of whale hearing. *Nature* 430:776–8.
- Nummela S, Wägar T, Hemilä S, Reuter T. 1999. Scaling of the cetacean middle ear. *Hear Res* 133:71–81.
- Nuttall AL. 1974. Tympanic muscle effects on middle-ear transfer characteristic. *J Acoust Soc Am* 56:1239–47.
- O'Connell CE, Arnason BT, Hart LA. 1997. Seismic transmission of elephant vocalizations and movement. *J Acoust Soc Am* 102:3124.
- O'Connell-Rodwell CE, Arnason BT, Hart LA. 2000. Seismic properties of Asian elephant (*Elephas maximus*) vocalizations and locomotion. *J Acoust Soc Am* 108:3066–72.
- O'Connell-Rodwell CE, Hart LA, Arnason BT. 2001. Exploring the potential use of seismic waves as a communication channel by elephants and other large mammals. *Am Zool* 41:1157–70.
- Oeckinghaus H, Schwartzkopff J. 1983. Electrical and acoustical activation of the middle ear muscle in a songbird. *J Comp Physiol* 150:61–7.
- Okanoya K, Dooling RJ. 1987. Hearing in passerine and psittacine birds: a comparative study of absolute and masked auditory thresholds. *J Comp Psychol* 101:7–15.
- Okanoya K, Dooling RJ. 1988. Hearing in the swamp sparrow, *Melospiza georgiana*, and the song sparrow, *Melospiza melodia*. *Anim Behav* 36:726–32.
- Partridge L, French V. 1996. Thermal evolution of ectotherm body size: why get big in the cold. In: Johnston IA, Bennett AF, editors. *Animals and temperature: Phenotypic and evolutionary adaptation*. Cambridge: Cambridge University Press. p. 265–92.
- Penna M, Márquez R. 2007. Amplification and spectral modification of incoming vocalizations inside burrows of the frog *Eusophus roseus* (Leptodactylidae). *Bioacoustics* 16:245–59.
- Penna M, Solís R. 1996. Influence of burrow acoustics on sound reception by frogs *Eusophus* (Leptodactylidae). *Anim Behav* 51:255–63.
- Pfennig KS. 2000. Female spadefoot toads compromise on mate quality to ensure conspecific matings. *Behav Ecol* 11:220–7.

- Pfennig KS, Ryan MJ. 2007. Character displacement and the evolution of mate choice: an artificial neural network approach. *Philos Trans R Soc Lond B Biol Sci* 362:411–9.
- Pollak G, Henson OW, Novick A. 1972. Cochlear microphonic audiograms in the “pure tone” bat *Chilonycteris parnellii parnellii*. *Science* 176:66–8.
- Reuter T, Nummela S, Hemila S. 1998. Elephant hearing. *J Acoust Soc Am* 104:1122–3.
- Richards DG, Wiley RH. 1980. Reverberations and amplitude fluctuations in the propagation of sound in a forest: implications for animal communication. *Am Nat* 115:381–99.
- Richardson WJ, Greene CR Jr, Malme C, Thomson DH. 1995. Marine mammals and noise. San Diego: Academic Press.
- Robertson JGM. 1986. Female choice, male strategies and the role of vocalizations in the Australian frog *Uperoleia rugosa*. *Anim Behav* 34:773–84.
- Roeder KD. 1962. The behavior of free flying moths in the presence of artificial ultrasonic pulses. *Anim Behav* 10:300–4.
- Roeder KD. 1966. Acoustic sensitivity of the noctuid tympanic organ and its range for the cries of bats. *J Insect Physiol* 12:843–59.
- Rosowski JJ. 1994. Outer and middle ears. In: Fay RR, Popper AN, editors. *Comparative hearing: mammals*. New York: Springer. p. 172–248.
- Rosowski JJ, Graybeal A. 1991. What did *Morganucodon* hear? *Zool J Linn Soc* 101:131–68.
- Ross RJ, Smith JJB. 1978. Detection of substrate vibrations by salamanders: inner ear sense organ activity. *Can J Zool* 56:1156–62.
- Ross RJ, Smith JJB. 1979. Detection of substrate vibrations by salamanders: eighth cranial nerve activity. *Can J Zool* 57:368–74.
- Ryan MJ. 1980. Female mate choice in a neotropical frog. *Science* 209:523–5.
- Ryan MJ. 1990. Signals, species, and sexual selection. *Am Sci* 78:46–52.
- Ryan MJ, Perrill SA, Wilczynski W. 1992. Auditory tuning and call frequency predict population-based mating preferences in the cricket frog, *Acris crepitans*. *Am Nat* 139:1370–83.
- Ryan MJ, Rand AS. 1990. The sensory basis of sexual selection for complex calls in the tungara frog, *Physalaemus pustulosus* (sexual selection for sensory exploitation). *Evolution* 44:305–14.
- Ryan MJ, Tuttle MD, Barclay RMR. 1983. Behavioral responses of the frog-eating bat, *Trachops cirrhosus*, to sonic frequencies. *J Comp Physiol* 150:413–8.
- Ryan MJ, Wilczynski W. 1988. Coevolution of sender and receiver: effect on local mate preference in cricket frogs. *Science* 240:1786–8.
- Spangler HG. 1988. Moth hearing, defense, and communication. *Annu Rev Entomol* 33:59–81.
- Stenfelt S, Hato N, Goode RL. 2002. Factors contributing to bone conduction: The middle ear. *J Acoust Soc Am* 111:947–59.
- Suga N, Simmons JA, Shimozawa T. 1974. Neurophysiological studies on echolocation systems in awake bats producing CF-FM orientation sounds. *J Exp Biol* 61:379–99.
- Tasaki I, Davis H, Eldredge DH. 1954. Exploration of cochlear potentials in guinea pig with a microelectrode. *J Acoust Soc Am* 26:765–73.
- ter Hofstede H, Ratcliffe J, Fullard J. 2008. The effectiveness of katydid (*Neoconocephalus ensiger*) song cessation as antipredator defence against the gleaning bat *Myotis septentrionalis*. *Behav Ecol Sociobiol* 63:217–26.
- Todd N, Rosengren SM, Colebatch JG. 2008. Tuning and sensitivity of the human vestibular system to low-frequency vibration. *Neurosci Lett* 444:36–41.
- van Dijk T. 1973. A comparative study of hearing in owls of the family Strigidae. *Neth J Zool* 23:131–67.
- Varanasi U, Malins DC. 1971. Unique lipids of the porpoise (*Tursiops gilli*): differences in triacyl glycerols and wax esters of acoustic (mandibular canal and melon) and blubber tissues. *Biochim Biophys Acta* 231:415–8.
- Vlaming MSMG, Feenstra L. 1986. Studies on the mechanics of the normal human middle ear. *Clin Otolaryngol* 11:353–63.
- von Békésy G. 1960. *Experiments in hearing*. New York: McGraw-Hill.
- Voss SE, Rosowski JJ, Merchant SN, Peake WT. 2000. Acoustic responses of the human middle ear. *Hear Res* 150:43–69.
- Walkowiak W. 1980. Sensitivity, range and temperature dependence of hearing in the grass frog and fire-bellied toad. *Behav Process* 5:363–72.
- Webster DB, Fay RR, Popper AN, editors. 1992. *The evolutionary biology of hearing*. New York: Springer.
- Werner YL, Igic PG. 2002. The middle ear of gekkonoid lizards: interspecific variation of structure in relation to body size and to auditory sensitivity. *Hear Res* 167:33–45.
- Werner YL, Igic PG, Seifan M, Saunders JC. 2002. Effects of age and size in the ears of gekkonoid lizards: middle-ear sensitivity. *J Exp Biol* 205:3215–23.
- Werner YL, Montgomery LG, Safford SD, Igic PG, Saunders JC. 1998. How body size affects middle-ear structure and function and auditory sensitivity in gekkonoid lizards. *J Exp Biol* 201:487–502.
- Wever EG, Bray CW. 1930. Action currents in the auditory nerve in response to acoustical stimulation. *Proc Natl Acad Sci USA* 16:344–50.
- Wever EG, Bray CW. 1936. Hearing in the pigeon as studied by the electrical responses of the inner ear. *J Comp Psychol* 22:353–63.
- Wilczynski W, McClelland BE, Rand AS. 1993. Acoustic, auditory, and morphological divergence in three species of neotropical frog. *J Comp Physiol* 172:425–38.

- Wilczynski W, Rand AS, Ryan MJ. 2001. Evolution of calls and auditory tuning in the *Physalaemus pustulosus* species group. *Brain Behav Evol* 58:137–51.
- Wilczynski W, Resler C, Capranica RR. 1987. Tympanic and extratympanic sound transmission in the leopard frog. *J Comp Physiol* 161:659–69.
- Willi UB, Bronner GN, Narins PM. 2006a. Middle ear dynamics in response to seismic stimuli in the Cape golden mole (*Chrysochloris asiatica*). *J Exp Biol* 209:302–13.
- Willi UB, Bronner GN, Narins PM. 2006b. Ossicular differentiation of airborne and seismic stimuli in the Cape golden mole (*Chrysochloris asiatica*). *J Comp Physiol* 192:267–77.
- Yu X, Lewis ER, Feld D. 1991. Seismic and auditory tuning curves from bullfrog saccular and amphibian papillar axons. *J Comp Physiol* 169:241–8.